

DTP, DCTD TUMOR REPOSITORY

A CATALOG OF IN VITRO CELL LINES, TRANSPLANTABLE
ANIMAL AND HUMAN TUMORS AND MICROARRAYS

**Operated by Charles River Laboratories, Inc.
under contract to the Biological Testing Branch for
The National Cancer Institute at Frederick
Frederick, Maryland 21702-1201**

Sponsored by:

Biological Testing Branch
Developmental Therapeutics Program
Division of Cancer Treatment and Diagnosis
National Cancer Institute
National Institutes of Health

www.dtp.nci.nih.gov

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DCTD Tumor Repository

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INTRODUCTION

The Division of Cancer Treatment and Diagnosis (DCTD), National Cancer Institute, has maintained since the early 1960s a low temperature repository of transplantable *in vivo*-derived tumors and *in vitro*-established tumor cell lines from various species. Currently located at the National Cancer Institute at Frederick in Frederick, Maryland, the DCTD Tumor Repository* serves as a resource for viable, contaminant-free experimental tumor lines, many of which are not obtainable elsewhere. The Repository makes these materials available to qualified investigators as a service to the research community.

The Repository's tumor collection contains a wide variety of frozen types of human and animal origin. Virtually all of the human tumors are xenografts grown in athymic nude mice, although there are some that grow in conditioned rats or in hamster's cheek pouch. Several mouse leukemia lines in the collection are resistant to single drugs of varying modes of action. Multidrug-resistant lines are also available. In addition, the collection includes variant sublines of B16 melanomas that exhibit a different degree of metastasis to various organs.

The tumors in this catalog are categorized by species, namely, human, hamster, guinea pig, mouse, rabbit and rat. Within animal species, the list is in alphabetical order by tumor designation. Tumors with numeric designations are listed at the end. Human tumors are grouped by tumor type.

We request that the DCTD Tumor Repository, National Cancer Institute at Frederick, Frederick, Maryland, be cited in publications as the source of tumor materials. We also request that reprints of publications be furnished to the Repository.

*Previous contract locations of the Tumor Repository include Microbiological Associates, Inc., Bethesda, MD; Arthur D. Little, Inc., Cambridge, MA; and Mason Research Institute, Worcester, MA.

ORDERING PROCEDURES

Tumor Fragments, Cell Lines and Yeast DTP, DCTD, NCI Repository Ordering Procedures and Material Transfer Agreement-A

Tumor materials are furnished to qualified investigators affiliated with recognized research laboratories *only*. **Items required to complete an order are:**

- a. **Original letter of request**, typed on official Institutional Letterhead
- b. **Purchase Order** or indicate payment by Visa, MasterCard, American Express, P.O. #, wire transfer, etc.
- c. **Two Material Transfer Agreements (MTA's)**, each with **original signatures from both the requestor and authorizing official** at your institution (required by NCI).

Request will be processed only after receiving the above three items. Faxed MTA's cannot be accepted.
Return all completed paperwork **via Mail or Courier** to:

Ms. Katherine Gill	
Biological Testing Branch	E: gillk@mail.nih.gov
DTP, DCTD, NCI-Frederick	T: 301-846-5483
Fairview Center, Suite 205	F: 301-846-6183
1003 West 7th Street	
Frederick, MD 21701-8531	

The letter should specify the material desired and should include a summary statement concerning the objectives of the studies in which the requested materials will be used. Completed MTA's for the material should be enclosed with the letter, unless a current MTA is already on file for the same item(s). **A method of payment must be indicated in the letter or attached to the paperwork.**

MTA's are valid for a period of three years. Newly requested material will require completion of a separate MTA. Re-orders by the same investigator for previously received material should indicate the date and MTA number of the earlier MTA in a letter of request.

Multiple cell lines, tumor lines, yeast strains and microarrays can be ordered using a single MTA. Each must be clearly identified under #1 on the MTA or through use of an addendum.

The following fees include shipping in those cases where standard delivery methods can be used:

	<u>NCI/NIH Investigators</u>	<u>Academia Domestic & Canadian</u>	<u>Non-Academic Domestic and International</u>
Cell or Tumor Lines:			
Each individual frozen vial	\$75.00	\$150.00	\$200.00
NCI anti-cancer cell line panel (when ordering complete panel 59 lines)	\$3,350.00	\$6,700.00	\$8,850.00
	<u>Academia</u>	<u>Non-Academic Domestic and International</u>	
Yeast Strains:			
One strain	\$150.00	\$200.00	
Complete set (16 strains)	\$1,800.00	\$2,400.00	

International Recipients: The DCTD Tumor Repository obtains United States Exportation Declarations for each shipment. **International recipients must obtain the applicable import permits as required by the recipient's country.** Send a copy with the MTA or state the Import number in your letter of request. Retaining a Broker to get your shipment through Customs is strongly suggested. In cases where standard shipping methods are not applicable, the recipient will be required to cover all excess charges associated with shipping. Please contact NCI in advance to determine your eligibility for standard shipping rates. Material will be sent freight prepaid.

Inquiries regarding material availability and invoicing should be directed to:

Jane Shelton T: 301 846-5025 F: (301) 846-6941 E: shelton@dpax2.ncifcrf.gov

Shipping questions should be directed to:

Vicky Clark T: 301 846-5025 F: 301 846-6941 E: clarkv@dpax2.ncifcrf.gov

Scientific questions should be directed to:

Dr. Melinda Hollingshead T: (301) 846-5608 F: (301) 846-6183 E: hollingm@mail.nih.gov

DTP, DCTD, NCI REPOSITORY
Cell Lines, Fragments, Yeast and TARP Microarrays

REMIT TO ADDRESS :
Charles River Laboratories, Inc.
GPO Box 27812
New York, NY 10087-7812

FEDERAL TAX ID # 76-0509980

Charles River Laboratories accepts the following credit cards for remittance: VISA, MASTERCARD and AMERICAN EXPRESS. Purchase orders, purchase order numbers and checks are also accepted.

For W-9 information and invoicing questions please contact:

Ms. Jane Shelton at shelton@dtpax2.ncifcrf.gov , Phone: 301-846-5025 or Fax: 301-846-6941

INTERNATIONAL:

ELECTRONIC FUNDS WIRE TRANSFER INFORMATION EFT/ACH WIRES

JP Morgan
New York, NY
ABA Number: 021000021
Account of: Charles River Laboratories Lock Box Account
DDA (Checking) Account Number: 304-264466
Swift Number: CHASUS33
ACH HELP DESK: (800) 447-3593
WIRE HELP DESK: (866) 223-0359

*** Please note: JPM uses the same ABA Number for both wires and ACH Payments
CTX Format required to utilize EFT/ACH*

The full invoice amount is to be paid. Any Bank wire fees incurred are at the purchaser's expense.

National Cancer Institute

MATERIAL TRANSFER AGREEMENT-A
Cell lines maintained in the NCI-DCTD Repository

This Material Transfer Agreement ("MTA") has been adopted for use by the National Cancer Institute ("NCI") for transfers of cell lines from the Division of Cancer Treatment and Diagnosis ("DCTD") Tumor Repository ("Research Material").

Recipient: _____
Name of Recipient Investigator and Recipient Institution

1. NCI agrees to transfer to Recipient named above the following Research Material:

(use an attachment page if necessary)

2. THIS RESEARCH MATERIAL MAY NOT BE USED IN HUMAN SUBJECTS. The Research Material will only be used for research purposes by Recipient's investigator in his/her laboratory, for the research project described below, under suitable containment conditions.

3. This Research Material will be used by Recipient's investigator solely in connection with the following research project ("Research Project") described with specificity as follows (use an attachment page if necessary):

4. In all oral presentations or written publications concerning the Research Project, Recipient will acknowledge NCI's contribution of this Research Material unless requested otherwise. To the extent permitted by law, Recipient agrees to treat in confidence, for a period of three (3) years from the date of its disclosure, any of NCI's written information about this Research Material that is stamped "CONFIDENTIAL,"

5. This Research Material represents a significant investment on the part of NCI. Recipient's investigator therefore agrees to retain control over this Research Material and further agrees not to transfer the Research Material to other people not under her or his direct supervision without advance written approval of NCI.

6. This Research Material is provided as a service to the research community. IT IS BEING SUPPLIED TO RECIPIENT WITH NO WARRANTIES, EXPRESS OR IMPLIED, INCLUDING ANY WARRANTY OF MERCHANTABILITY OR FITNESS FOR A PARTICULAR PURPOSE. NCI makes no representations that the use of the Research Material will not infringe any patent or proprietary rights of third parties.

7. Recipient may retain title to the patent rights in inventions made by its employees in the course of the Research Project. Recipient agrees not to claim, infer, or imply Governmental endorsement of the Research Project, the institution or personnel conducting the Research Project or any resulting product(s). Unless prohibited by law from doing so, recipient agrees to hold the United States Government harmless and to indemnify the Government for all liabilities, demands, damages, expenses and losses arising out of Recipient's use for any purpose of the Research Material.

8. The undersigned Recipient expressly certifies and affirms that the contents of any statements made herein are truthful and accurate.

9. This MTA shall be construed in accordance with Federal law as applied by the Federal courts in the District of Columbia.

Date: _____

Recipient Investigator's Signature and Title

Date: _____

Authorized Signature and Title, for Recipient's Institution (**Note:** Authorized Signature the authority to bind the Institution to the terms of this Material Transfer Agreement).

→ Recipient's Shipping Address:

→ Recipient's Phone, Fax, Email:

Authorized Signature for NCI: _____ Date: _____
Melinda G. Hollingshead, D.V.M., Ph.D., Chief, BTB, DTP, DCTD, NCI-Frederick

NCI's Mailing Address:

Ms. Katherine Gill
Biological Testing Branch
DTP/DCTD/NCI
1003 W. Seventh Street
Fairview Center, Suite 205
Frederick, MD USA 21701-8531

Phone: 301.846.5483
Fax: 301.846.6183
Email: gillk@mail.nih.gov

SHIPMENT OF TUMORS

The transplantable tumors are distributed as frozen vials of tumor tissues or cell suspension. Transplantable tumors as well as cell culture lines are shipped as frozen vials of tissues on dry ice. Each tumor shipment includes an information sheet showing, among other items, the proper tumor designation, cryopreserved date, *in vivo* host, etc.

Requested tumors are shipped two to three weeks after receipt of all completed paperwork. Domestic shipments leave the Repository no later than Wednesday in order to reach their destination on week days. Foreign shipments are sent according to final destination. Before shipment, the Repository notifies the Recipient of the waybill number and carrier information by e-mail or fax. Recipient must notify the Repository or make arrangements for receipt of the tumor lines, in the event that they will not be available to receive the shipment. An invoice for payment will follow and payment is due upon receipt.

SUBMISSION OF TUMORS FOR CRYOPRESERVATION

Investigators who have unique and novel experimental tumor lines and wish to submit their tumors to the Repository for cryopreservation and storage should write a letter of intent to the Project Officer. Upon acceptance, the Project Officer will inform the investigator in writing, and will provide instructions on the procedure for shipping tumor materials to the Repository. Tumor tissues or cells (frozen or ambient) are preferred over tumor-bearing animals.

At the Repository, the tumor line(s) will be tested for viral and bacterial contamination. When proven "clean," the line(s) will be expanded, *in vivo* or *in vitro* as appropriate, for large batch cryopreservation. Viability and growth of the frozen tumors will be evaluated. The tumors will be included in the Repository's inventory, and upon joint approval of the submitting investigator and the Project Officer, they will be made available for distribution to the scientific community.

MOUSE TUMORS FROM THE JACKSON LABORATORY

These tumors formerly were maintained and distributed by the Jackson Laboratory. The list of available tumors begins on page 39 of the DCTD Tumor Repository Catalog. They were cryopreserved at EG&G Mason Research Institute and are distributed only as vials of frozen tumor tissue. The required host animals for carrying the JAX tumors in serial transplantation may be obtained from:

Animal Resources
The Jackson Laboratory
600 Main Street
Bar Harbor, ME 04609
USA
T: 800.422.MICE
207.288.5845
F: 207.288.6150

FREEZING PROCEDURE

Solid tumors are frozen as 2 x 2 x 2 mm fragments suspended in a freezing medium. Ascites or tissue culture cells are frozen as single cell suspension at a concentration of 10^6 - 10^7 cells per ml. The freezing medium consists of appropriate tissue culture growth medium plus 10% DMSO and 10% fetal bovine serum.

Aseptically harvested ascites tumors are diluted in the freezing medium at a concentration of 10^6 - 10^7 cells per ml. One ml suspension is pipetted into each 2 ml vial (Nunc cryotube). The vials are screw-capped tightly and labeled with Repository number. Tissue culture cells are prepared in a similar manner. For solid tumors, the aseptically excised tumor tissue is cut into 2 x 2 x 2 mm fragments after freeing it of necrotic materials. The fragments are placed in vials containing 1.5 ml of freezing medium.

The processed tumors are frozen initially in a controlled slow-rate freezing apparatus at the rate of 0.5°C per minute to -20°C and 1°C per minute to -80°C. The frozen vials are stored in specific locations in the liquid nitrogen freezers in the Repository after the controlled freezing cycle.

RECOMMENDED THAWING PROCEDURE

Frozen tumor cells or tissues received from the Repository should be kept frozen at -70°C or lower until ready for use. For prolonged storage (more than two days), liquid nitrogen freezers are recommended.

The vials in which the cell lines are stored are reliable; however, they are very susceptible to contamination if thawed in a contaminated water bath. Thawing should be rapid, i.e., within 60-90 seconds. Place the vial in a warm water bath at 37-40°C and agitate vigorously to thaw. Immerse the vial in 70% ethanol before uncapping. Implant immediately after thawing. The concentration of DMSO is not toxic to tissues and implantation may be made directly from the vial.

For tissue culture, transfer the contents of the vial into a petri dish or a flask containing at least 10 volumes of the recommended culture medium and incubate. In order to remove the protective freezing additive (DMSO) from the culture medium, we suggest that the culture medium be changed 24 hours after seeding. If it is desired that DMSO be removed immediately, centrifuge the diluted suspension at approximately 125 x g for 10 minutes, discard the supernatant, and resuspend the cells in an appropriate volume of growth medium.

CAUTION: We strongly recommend wearing protective glasses or face shields when thawing tissues in glass vials.

TUMOR TRANSPLANTATION: It is recognized that transplantable tumor systems are experimental tools for investigators in scientific disciplines other than tumor biology or transplantation immunogenetics. Therefore, we encourage investigators with limited transplantation experience to contact the Tumor Repository for more detailed information on techniques.

BACTERIAL AND VIRAL MONITORING OF CRYOPRESERVED TISSUES

In addition to testing all freeze-runs for bacterial contamination, cryopreserved tissues are tested for viral contamination by the MAP test. The viruses tested for are as follows: pneumonia virus of mice (PVM), reo virus-type 3 (Reo 3), Theiler's virus, murine encephalitis (GD VII), polyoma (Poly), minute virus of mice (MVM), mouse hepatitis virus (MHV), lymphocytic choriomeningitis (LCM), Sendia virus (Send), ectromelia, mouse pox (Ectro), lactic dehydrogenase virus (LDH).

IN VITRO ESTABLISHED CELL LINES:

- A. **Quality Control and Characterization** - The quality control and characterization procedures for the incorporation of new cell lines into the Tumor Repository are as follows: Upon receipt, each cell line is immediately transferred to fresh antibiotic-free medium and cultured for one week, after which it is tested for mycoplasma (PPLO) contamination. Standard culture procedures under aerobic and anaerobic conditions, as well as the orcein staining procedure of Fogh, are used. The PPLO medium is extremely rich, and this procedure will also detect most bacterial and fungal contaminants.
- B. **Freezing and Storage** - The cell cultures are frozen in ampules containing 1.0 ml of cell suspension at $2-6 \times 10^6$ cells/ml in fresh culture medium containing 10% DMSO. Freezing is performed as on page 6. Twenty-four hours after freezing, a representative ampule is removed, thawed, and viable cell count is performed, using the trypan blue dye exclusion procedure. The culture is also tested for its ability to initiate a heavy viable culture. Cell preparations which show less than 50% viability or poor growth are discarded and a new lot is prepared.
- C. **Recommended Procedure for Thawing Frozen Cell Cultures** - The vials in which the cell lines are stored are reliable; however, they are very susceptible to contamination if thawed in a contaminated water bath. The following procedures are provided to eliminate this problem.

Remove the ampule from the dry ice container and place it directly into a 37-40°C water bath (or vessel) of freshly drawn water containing an effective concentration of disinfectant. The thawing should be vigorous and rapid (within 40-60 seconds). As soon as the thawing is complete, remove the ampule from the water bath and immerse in 70% ethanol at room temperature. All of the operations from this point should be carried out under strict aseptic conditions in a sterile room, cubicle, or hood.

Transfer the contents of the ampule (1 ml volume) into a 100 mm petri dish or 25 cm² flask containing 8-10 ml of the recommended culture medium, and incubate

at the appropriate temperature and carbon dioxide level.

In order to remove the protective freezing additive (DMSO) from the culture medium, we suggest that the culture medium be changed 24 hours after thawing. If it is desired that the freezing additive be removed immediately or that a more concentrated cell suspension be obtained, centrifuge the above diluted suspension at approximately 125 x g for 10 minutes, discard the fluid, and resuspend the cells in an appropriate volume of growth medium.

HUMAN TUMORS

SPECIES: HUMAN			
Tumor Designation	Histologic Type	General Information	Species and/or Strain of Transplantability
BREAST			
COO-G	Mammary Carcinoma	Primary explant established <i>in vivo</i> in athymic nude mice by Dr. B. Giovanella, Stehlin Foundation.	Nude Athymic Mice
DU4475	Mammary Carcinoma	Primary explant from cutaneous tumor nodule in region of mastectomy established <i>in vitro</i> by Dr. A.J. Langlois, Duke University Medical Center; then adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
ELL-G	Mammary Carcinoma	Primary explant established <i>in vivo</i> in athymic nude mice by Dr. B. Giovanella, Stehlin Foundation.	Nude Athymic Mice
HIG-G	Mammary Carcinoma	Primary explant established <i>in vivo</i> in athymic nude mice by Dr. B. Giovanella, Stehlin Foundation.	Nude Athymic Mice
MCF/7	Mammary Carcinoma	Primary explant from pleural effusate established <i>in vitro</i> by Dr. H.D. Soule, Michigan Cancer Foundation; then adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MDA-MB-436	Mammary Carcinoma	Primary explant from pleural effusate established <i>in vitro</i> by Dr. Relda Cailleau, M.D. Anderson Hospital and Tumor Institute; then adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MX-1	Mammary Carcinoma	Primary xenotransplant from an infiltrating duct carcinoma (CLO-G). Adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella.	Nude Athymic Mice
MX-2	Mammary Carcinoma	Adapted to <i>in vivo</i> from <i>in vitro</i> established pleural effusate.	Nude Athymic Mice
SW-613	Mammary Carcinoma	Established from <i>in vitro</i> line.	Nude Athymic Mice
VAN-G	Mammary Carcinoma	Primary explant established <i>in vivo</i> in athymic nude mice by Dr. B. Giovanella, Stehlin Foundation.	Nude Athymic Mice
LUNG			
LX-1	Lung, undifferentiated carcinoma	Xenotransplant from a metastasis to subcutaneous tissue (DOY-G). The primary lung tumor was an oat cell carcinoma. Adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella.	Nude Athymic Mice
COS-G	Lung, papillary carcinoma	Xenotransplant adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella.	Nude Athymic Mice

SPECIES: HUMAN			
Tumor Designation	Histologic Type	General Information	Species and/or Strain of Transplantability
LUNG (continued)			
H-MESO-1	Lung, mesothelioma	Xenotransplant from a primary tumor received from Dr. R.M. Williams and Dr. A. Rossini. Adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
H-MESO-1A	Lung, mesothelioma	H-MESO-1 converted to ascites form by Dr. A.E. Bogden.	Nude Athymic Mice
NCI-H23 H23	Lung, nonsmall cell, adenocarcinoma	Obtained from Dr. Adi Gazdar.	Nude Athymic Mice
NCI-H460 H460	Lung, nonsmall cell, epid.	Obtained from Dr. Adi Gazdar.	Nude Athymic Mice
MRI-H-165	Lung, squamous cell carcinoma	Primary xenotransplant adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MRI-H-266	Lung, poorly differentiated carcinoma	Xenotransplant from a metastasis, adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
COLON			
CX-1 (HT-29)	Colon, adenocarcinoma	Primary, untreated adenocarcinoma (HT-29) adapted to <i>in vitro</i> cell culture by Dr. Jergen Fogh; then adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden. Secretes CEA.	Nude Athymic Mice
CX-2	Colon, carcinoma	Primary xenotransplant (CA-1) adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella. Received at MRI from Dr. A. Ovejera.	Nude Athymic Mice
CX-3	Colon, carcinoma		Nude Athymic Mice
CX-5	Colon, adenocarcinoma	Xenotransplant from an untreated metastasis (SQU-G) adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella.	Nude Athymic Mice
GOB-G	Colon, adenocarcinoma	Xenotransplant adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella.	Nude Athymic Mice
HCC-2998	Colorectal carcinoma	Obtained from Dr. I.J. Fidler.	Nude Athymic Mice
HCT-15	Colon, carcinoma	Established from <i>in vitro</i> line.	Nude Athymic Mice
KLO-G	Colon, adenocarcinoma	Xenotransplant adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella.	Nude Athymic Mice
KM20L2	Colon, adenocarcinoma	Obtained from Dr. I.J. Fidler.	Nude Athymic Mice
MRI-H-194	Colon, adenocarcinoma	Xenotransplant from a metastasis adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
LOVO I	Colon, adenocarcinoma	Established from <i>in vitro</i> line.	Nude Athymic Mice

SPECIES: HUMAN			
Tumor Designation	Histologic Type	General Information	Species and/or Strain of Transplantability
COLON (continued)			
LOVO II	Colon, adenocarcinoma	Established from <i>in vitro</i> line.	Nude Athymic Mice
MRI-H-250	Colon, carcinoma	Xenotransplant from a metastasis adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MELANOMA			
BOW-G	Melanosarcoma	Xenotransplant from metastasis adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella.	Nude Athymic Mice
DEAC-7	Melanoma	Received as cryopreserved ampules from Dr. S. Warren and Dr. W.B. Patterson.	Hamster Cheek Pouch
FO #1	Melanoblastoma	Adapted to <i>in vivo</i> transplantation (FOS-G) by Dr. B. Giovanella.	Nude Athymic Mice
NIS-G	Melanosarcoma	Xenotransplant adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella.	Nude Athymic Mice
TRI-G	Melanoma	Xenotransplant adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella.	Nude Athymic Mice
WIL-G	Melanoma	Xenotransplant adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella.	Nude Athymic Mice
MRI-H-121B	Melanoma, malignant	Primary xenotransplant adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MRI-H-187	Melanoma, epithelioid, melanotic	Xenotransplant from metastasis adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MRI-H-221	Melanoma, malignant	Xenotransplant from metastasis adapted to <i>In vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MRI-H-255	Melanoma	Xenotransplant adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
CERVIX			
MRI-H-130	Cervix, squamous cell carcinoma	Primary xenotransplant adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MRI-H-177	Cervix, squamous cell carcinoma	Xenotransplant from a metastasis adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MRI-H-186	Cervix, invasive, large cell, nonkeratinizing, squamous cell carcinoma	Primary xenotransplant adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MRI-H-196	Cervix, poorly differentiated squamous cell carcinoma	Primary xenotransplant adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice

SPECIES: HUMAN			
Tumor Designation	Histologic Type	General Information	Species and/or Strain of Transplantability
CERVIX (continued)			
MRI-H-215	Cervix, invasive, large cell, nonkeratinizing, poorly differentiated, epidermoid carcinoma	Primary xenotransplant adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
KIDNEY			
MRI-H-121	Kidney, carcinoma	Xenotransplant from a metastasis adapted to <i>in vivo</i> transplant by Dr. A.E. Bogden.	Nude Athymic Mice
MRI-H-166	Kidney, transitional cell carcinoma	Primary xenotransplant adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
ENDOMETRIUM			
MRI-H-147	Endometrium, carcinoma, Müllerian duct	Primary xenotransplant adapted <i>in vivo</i> trans-plantation by Dr. A.E. Bogden.	Nude Athymic Mice
MRI-H-171	Endometrium, carcinoma	Primary xenotransplant adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MRI-H-220	Endometrium, carcinoma	Primary xenotransplant adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MRI-H-253	Endometrium, carcinoma	Primary xenotransplant adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
OVARY			
MRI-H-207	Ovary, undifferentiated carcinoma	Primary xenotransplant adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MRI-H-227	Ovarian Adenocarcinoma	Primary explant established <i>in vivo</i> by Dr. A.E. Bogden. Received from Dr. R. Hunter, University of Massachusetts Medical School.	Nude Athymic Mice
MRI-H-258	Ovarian Adenocarcinoma	Primary explant established <i>in vivo</i> by Dr. A.E. Bogden. Received from Dr. R. Hunter, University of Massachusetts Medical School.	Nude Athymic Mice
MRI-H-273	Ovarian Carcinoma	Originated from metastasis. Established <i>in vivo</i> by Dr. A.E. Bogden. Received from New England Deaconess Hospital.	Nude Athymic Mice
MRI-H-1834	Ovarian Carcinoma	Primary explant established <i>in vivo</i> by Dr. A.E. Bogden. Received from Dr. R. Hunter, University of Massachusetts Medical School.	Nude Athymic Mice
SWA-G	Ovarian Carcinoma	Xenotransplant adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella.	Nude Athymic Mice

SPECIES: HUMAN			
Tumor Designation	Histologic Type	General Information	Species and/or Strain of Transplantability
SARCOMA			
HS-1	Sarcoma	No historical information available.	Conditioned Rats
OGL-G	Sarcoma, spindle cell, periosteal osteogenic	Primary xenotransplant adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella.	Nude Athymic Mice
DEL-G	Sarcoma	Primary xenotransplant adapted to <i>in vivo</i> transplantation by Dr. B. Giovanella.	Nude Athymic Mice
EPIDERMOID			
HEP-3	Epidermoid carcinoma	No historical information available.	Conditioned Rats
DEAC-1	Mucoepidermoid carcinoma	Received as cryopreserved ampules from Dr. S. Warren and Dr. W.B. Patterson.	Hamster Cheek Pouch
CNS			
SF 295	Glioblastoma	Obtained from Dr. Rosenblum.	Nude Athymic Mice
MISCELLANEOUS			
CWR-22	Prostate, adenocarcinoma	Received from Dr. T. Pretlow, <i>in vivo</i> cultivation requires testosterone supplementation. Not an <i>in vitro</i> cell line.	Nude Athymic Mice
DAU	Burkitt's lymphoma	Received from Dr. T. Griffin, adapted to <i>in vivo</i> transplantation by Dr. A.E. Bogden.	Nude Athymic Mice
MRI-H-254	Stomach, adenocarcinoma	No historical information available.	Nude Athymic Mice
MRI-H-1579	Prostate Adenocarcinoma	Primary explant established <i>in vivo</i> by Dr. A.E. Bogden. Received from Dr. Blute, St. Vincent Hospital, Worcester, MA.	Nude Athymic Mice

Note: *Human In Vitro Established Cell Lines are on pages 59-62.*

HAMSTER TUMORS

SPECIES: HAMSTER				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
Fibrosarcoma	Fibrosarcoma	Ascites	Not Specified	
H-12	Mesothelioma	Solid	Golden Syrian	
H-75	Mesothelioma	Solid	Golden Syrian	
Islet Cell	Pancreatic Adeno- carcinoma	Solid	Golden Syrian	
Lymphosarcoma	Lymphosarcoma	Ascites	Not Specified	
Melanoma	Melanotic Melanoma	Solid	Not Specified	
Pan #1 (Fortner)	Pancreatic Duct Adeno- carcinoma	Solid	Not Specified	
SB #1 (Fortner)	Small Bowel Adeno- carcinoma	Solid	Not Specified	
TG1-4	Mesothelioma	Solid	Golden Syrian	
TS1-4	Epidermoid Carcinoma	Solid	Golden Syrian	
10-24	Mesothelioma	Ascites	Golden Syrian	
1382J	Liver Carcinoma	Solid	Golden Syrian	
2309V	Pancreatic Islet β Cell Adenocarcinoma	Solid	Golden Syrian	
4671	Pancreatic Duct Adeno- carcinoma	Solid	Golden Syrian	Line B is insulin- secreting
6973P	Leiomyosarcoma	Solid	Golden Syrian	
8721R	Renal Carcinoma	Solid	Golden Syrian	
8746Q	Uterine Adeno- carcinoma	Solid	Golden Syrian	
9242e	Parotid Acinar Cell Adenocarcinoma	Solid	Golden Syrian	
10838	Seminoma	Solid	Golden Syrian	
11348P	Pulmonary Squamous Cell Carcinoma	Solid	Golden Syrian	
11963V	Leiomyosarcoma	Solid	Golden Syrian	
22047	Adenocarcinoma	Solid	Golden Syrian	

GUINEA PIG TUMORS

SPECIES: GUINEA PIG				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
Line 1	Hepatocarcinoma	Ascites	Strain 2	
Guinea Pig GP Line 10	Hepatocarcinoma	Ascites	Strain 2	
Nitrosamine-induced Hepatoma	Hepatoma	Solid	Strain 39	

MOUSE TUMORS

Two addenda have been inserted in this section to facilitate identification or selection of mouse tumors by histologic type (Addendum A) and by mouse strain (Addendum B). A third (Addendum C) is a list of drug-resistant murine leukemias together with treatment information.

ADDENDUM A: MOUSE TUMORS LISTED BY HISTOLOGICAL TYPE

Adrenal

AT
LAF₁

Anaplastic Carcinoma

dbrB (Jax)

Bladder

FCB

Colon

CA07/A
CA36/Ara C
CA51
Colon 26
Colon 38

Ependymoblastoma

Ependymoblastoma

Fibrosarcoma

36257 TTT
Hepatoma 129

38290 TTT
Hepatoma 134

46362 TTT

46363 TTT

FB SAR
SaD2 (Jax)

Melanoma

Glioma

Glioma 26
Glioma 261

Hemangioendothelioma

36230 TLT
42021 TCT
42022 TST
42052 TST
42076 TST
44316 TST
44347 TST

Hepatoma

BW 7756 (JAX)
H6 (JAX)
Hepatoma 129
Hepatoma 134

Inguinal

Krebs Ascites
Krebs-2

Leukemias

C58/J Spont.
C1498
Egg/Mouse
E Male/Gross
Gross
L1210
L4946
L5178Y
P288
P388
P815
P1534
P1798
RBL-n

Lung

ASB XIV
C4461
CAD2
LC-12
Lewis
M4898
Madison
Nettesheim

Lymphoma &
Lymphosarcoma

BL12
EL-4 Male
L18464
LSTRA
Mecca
NK
6C3HED

Mammary

Adenocarcinoma 755
BW10232 (Jax)
CE1460 MACA
CH
C3HBA (Jax)
DBA/2 Spont. M114
Ehrlich Ascites
EMT-6
Gross
H2712 (Jax)
Klein
MA13C
MA16C
MC-n
MCS-1
MXT
Spont. DBA/2

B16

ADDENDUM A (continued)

Harding-Passey
HP (Jax)

Nervous System

C1300 (Jax)
Ependymoblastoma
Glioma

Osteogenic

HE10734

Pancreas

PAN 02
PAN 03

A+T
BW8685 (Jax)
BW8883 (Jax)
T+T #15
T+T #97

Plasmacytoma

ADJ-PC-n
LPC-1
MOPC-n
MPC-n
RPC-n
YPC-1
70429

Reticulum Cell

Friend Virus Leukemia
M5076
SJL/JW
91632

Sarcoma

Lewis
MA387
METH-A
MS-2
S37
S180
Sa-1 (Jax)

Squamous

LC-12

Testicular

M5480

Teratosarcoma

LS402AX

Thymus

Reif-Allen

Pituitary

*Refer to Inventory for details. A designation followed by n, e.g., MOPC-n, indicates that there is a series of tumors with this main designation, e.g., MOPC-4, MOPC-17, MOPC-21, etc.

ADDENDUM B (continued)

ADDENDUM B: MOUSE TUMORS LISTED BY HOST STRAIN*

INBRED HOSTS

<u>A/HE</u>	P1798 <u>BALB/c (cont'd)</u>	BL12 BW 10232 (Jax)
C4461 Hauschka Ascites Klein (TA3)	RPC- <u>n</u> S37 YPC-1 1247 36257 TTT 38290 TTT 42021 TCT 44316 TST 44316 LTST 44347 TST 46363 TTT	<u>C57BL/6 (cont'd)</u> C1498 Colon 38 EL-4 E Male Gross Ependymoblastoma FCB Glioma 26 Glioma 261 L18464 Lewis Lung Lewis Sarcoma T241 LS402AX M5076 M5480 NK-Lymphoma PAN 02 PAN 03 RBL- <u>n</u> T+T #15 42022 TST 42052 TST 42076 TST 46362 TTT 91632
<u>A/J</u>	<u>CE</u> CE1460 MACA	<u>C57L/J</u> BW7756 (Jax) BW8883 (Jax)
C1300 (Jax) H6 (Jax) Sa 1 (Jax)	<u>C3H</u> C3HBA (Jax) FB SAR Gross Leukemia H2712 (Jax) HE 10734 Hepatoma 129 Hepatoma 134 J30237 Krebs Ascites Carcinoma Krebs 2 Carcinoma MA13C MA16C Mecca X5563 6C3HED 70429	<u>C57BR/cdj</u> BW8685 (Jax)
<u>AKR</u>	<u>C57BL/6</u> Adenocarcinoma 755 B16	<u>C58</u>
L4946 MA387 Mecca Reif-Allen		
<u>BALB/c</u>		
ADJ-PC- <u>n</u> ASB XIV CA07/A CA36/Ara C CA51 Colon 26 EMT-6 Harding-Passey HP (Jax) LC-12 LPC-1 LSTRA M4898 Madison Lung MC- <u>n</u> METH-A Moloney Sarcoma MOPC- <u>n</u> MPC- <u>n</u> MS-1 MS-2		

ADDENDUM B (continued)

C58J/Spont.

DBA/1

CaD1 (Jax)
dbrB (Jax)
S37 (Jax)
S91 (Jax)
T1703 (Jax)

DBA/2

CAD2
DBA/2 Spont. M114
Egg/Mouse Leukemia
Friend Virus Leukemia

DBA/2 (cont'd)

Gross Mammary
denocarcinoma
L1210
L5178Y
Nettesheim
P288
P329
P388
P815
P1534
S180

SaD2 (Jax)
DBA/2 (cont'd)

Spont. DBA/2 Mammary
T1699 (Jax)

SJL/J
SJL/JW

129
LS402AX

F₁ HYBRIDS AND NON-INBRED HOSTS

Swiss

Ehrlich Ascites
S180

BDF₁

MXT

Various resistant lines of
L1210 and P388

LAF₁

AT
LAF₁
MST
T+T #97

CAF₁

Lymphoma-2

CDF₁

R-n
RC-2
102A

Various resistant lines of
L1210 and P388

*Refer to Inventory for details. A designation followed by n, e.g., MOPC-n, indicates that there is a series of tumors with this main designation, e.g., MOPC-4, MOPC-17, MOPC-21, etc.

ADDENDUM C: DRUG-RESISTANT MURINE LEUKEMIAS AND TREATMENT INFORMATION

L1210 Lymphoid Leukemia

L1210/TSC (NSC-729)	L1210/Ara-C (NSC-63878)
L1210/MTX (NSC-740)	L1210/cis-DDP (NSC-119875)
L1210/6MP (NSC-755)	L1210/Anhydro Ara C (NSC-145668)
L1210/L-PAM (NSC-8806)	L1210/Ftorafur (NSC-148958)
L1210/NSC-19622	L1210/L-Alanosine (NSC-153353)
L1210/5FU (NSC-19893)	Note: Reo3 ⁺
L1210/CTX (NSC-26271)	L1210/BCNU (NSC-409962)
L1210/DF8 (NSC-29630)	L1210/C95 (NSC-740, 755, 26271)
L1210/HU (NSC-32065)	L1210/FR3 DCM/R 100a
L1210/MeGAG (NSC-32946)	L1210/FR8/DCM
L1210/NSC-38280	L1210/RT8 (Folate Reductase)
L1210/DTIC (45388)	L1210/M-773
L1210/TIC (NSC-60339)	L1210/6MP/6TG

*Treatment information, where available, is given in the following pages. When resistant lines are shipped, treatment information, if any, is included.

P388 Lymphocytic Leukemia

P388/MTX (NSC-740)	P388/L-Alanosine (NSC-153353)
P388/Actinomycin D (NSC-3053)	P388/Acivicin (NSC-163501)
P388/DON (NSC-7365)	P388/Amsacrine (NSC-249992)
P388/L-PAM (NSC-8806)	P388/Anthracenedione (NSC-287513)
P388/5-FU (NSC-19893)	Not
P388/CPA (NSC-26271)	e: Reo3 ⁺
P388/Ara C (NSC-63878)	P388/Mitoxantrone (NSC-299195 + 301739)
P388/Daunomycin (NSC-82151)	P388/Ara-A + 2'dcF (NSC-404241 + 218321)
P388/5-Azacytidine (NSC-102816)	P388/BCNU (NSC-409962)
P388/DDP (NSC-119875) Note: Reo3 ⁺	
P388/ADR (NSC-123127)	

Other Resistant Leukemias

L5178/L-Ase R
P288/MTX (NSC-740)
P815/VLB (NSC-49842)

DRUG-RESISTANT MURINE LEUKEMIAS - TREATMENT INFORMATION FOR LINES RECEIVED
FROM SOUTHERN RESEARCH INSTITUTE AND ARTHUR D. LITTLE, INC.

Tumor Line	Host of Origin, Resistant Line	Passage Inoculum (i.p.)	Treatment Used with Serial Passage				Optimal Treatment to Check Degree of Resistance			
			NSC #	mg/Kg	Route	Schedule	NSC #	mg/Kg	Route	Schedule
L1210/TSC (NSC-729)	DBA/2 or CDF1	10 ⁵	729	5.0	i.p.	Days 1-6	729	6.0	i.p.	Q3Hx8 Days 1,5,9
L1210/6-MP (NSC-755)	DBA/2	10 ⁶	NONE				755	50.0	i.p.	QD1-9 days
L1210/L-PAM (NSC-8806)	BDF1	10 ⁶	8806	7.5	i.p.	Day 2 only	8806	15.0	i.p.	Day 1 only
L1210/CPA (NSC-26271)	DBA/2	10 ⁵	NONE				26271	265.0	i.p.	Day 1 only
L1210/HU (NSC-32065)	DBA/2 or CDF1	10 ⁵	32065	130.0	i.p.	Days 1-6	32065	60.0	i.p.	Q3Hx8 Days 1,5,9
L1210/ARA-C (NSC-63878)	DBA/2 or hybrid	10 ⁵	NONE				135962	125.0	i.p.	Day 1 only
L1210/DDP (NSC-119875)	DBA/2 or CDF1	10 ⁶	119875	5.0	i.p.	Day 4 only	119875	8.0	i.p.	Day 1 only
L1210/BCNU (NSC-409962)	BDF1	10 ⁵	NONE				409962	30.0	i.p.	Day 1 only
P388/MTX (NSC-740)	DBA/2 or CDF1	10 ⁷	740	0.75	s.c.	Days 1-6	740	2.0	i.p.	QD1-9 days
P388/ACT-D (NSC-3053)	DBA/2 or CDF1	10 ⁷	3053	0.2	i.p.	Day 4 only	3053	0.5	i.p.	Day 1 only
P388/L-PAM (NSC-8806)	BDF1	10 ⁶	8806	7.5	i.p.	Day 2 only	8806	15.0	i.p.	Day 1 only
P388/5-FU (NSC-19893)	BDF1	10 ⁷	19893	20.0	s.c.	Days 1-6	19893	25.0	i.p.	QD1-9 days
P388/CPA (NSC-26271)	BDF1	10 ⁷	26271	100.0	s.c.	Day 4 only	26271	265.0	i.p.	Day 1 only
P388/VCR	BDF1	10 ⁷	NONE				67574	1.5	i.p.	Days 1,5,9

Tumor Line	Host of Origin, Resistant Line	Passage Inoculum (i.p.)	Treatment Used with Serial Passage				Optimal Treatment to Check Degree of Resistance			
			NSC #	mg/Kg	Route	Schedule	NSC #	mg/Kg	Route	Schedule
(NSC-67574)										
P388/AZACYT (NSC-102816)	DBA/2 or CDF1	10 ⁷	102816	40.0	i.p.	Day 4 only	102816	3.5	i.p.	QD1-9 days
P388/DDP (NSC-119875)	DBA/2 or CDF1	10 ⁷	119875	4.5	i.p.	Days 1&5	119875	5.3	i.p.	Days 1,5,9
P388/ADR (NSC-123127)	BDF1	10 ⁷	123127	6.0	i.p.	Day 2 only	123127	12.5	i.p.	Day 1 only
P388/ARA-A + 2'dcF (NSC-404241 + NSC-218321)	BDF1	10 ⁶	404241 + 218321	125.0 .02	i.p.	Days 2-4	404241 + 218321	60.0 0.05	i.p.	Q3Hx8, Days 1,5,9
							<u>OR</u>			
							404241 + 218321	150.0 0.25	i.p.	QD1-9 days
P388/BCNU (NSC-409962)	CDF1	10 ⁷	409962	25.0	i.p.	Day 2 only	409962	30.0	i.p.	Day 1 only

*NSC-218321 thirty minutes before NSC-404241 each time.

SPECIES: MOUSE				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
Adenocarcinoma 755 (CA755, Bagg-Jackson, Adenocarcinoma)	Mammary Adenocarcinoma	Solid, Ascites or Brei	C57BL	
ADJ-PC-6	Plasmacytoma	Solid or Ascites	BALB/c	
ASB XIV	Pulmonary Squamous Cell Carcinoma	Solid	BALB/c	MVM
AT (Clone Y ₁)	Adrenal	Solid	LAF ₁	
AtT/20	Anterior Pituitary	Solid	LAF ₁	
B16	Melanoma	Solid	C57BL/6	Also see Jax tumors
BL12 Sensitive	Lymphosarcoma	Solid	C57BL/Ka	
BL12/Hc Ra	Lymphosarcoma	Solid	C57BL/Ka	Resistant to cortisone
BW7756	Hepatoma	Solid	C57L/J	See Jax tumors
BW8685	Pituitary	Solid	C57BR/Cdj	See Jax tumors
BW8883	Pituitary	Solid	C57L/J	See Jax tumors
BW10232	Mammary Adenocarcinoma	Solid	C57BL/6J	See Jax tumors
C3HBA	Mammary Adenocarcinoma	Solid	C3H/An	See Jax tumors
C58/J Spontaneous	Leukemia	Spleen Brei	C58	
C95 Spleen/R				See L1210/C95
C1498	Myelogenous Leukemia	Solid	C57BL/6	
C4461	Lung Adenocarcinoma	Solid	A/He	
CA07/A	Colon Adenocarcinoma	Solid	BALB/c	
CA36/Ara C (NSC-63878)	Colon Adenocarcinoma	Solid	BALB/c	
CA51	Colon Adenocarcinoma	Solid	BALB/c	
CaD1	Mammary Adenocarcinoma	Solid	DBA/1J	See Jax tumors
CaD2	Mammary Adenocarcinoma	Solid	DBA/2	

SPECIES: MOUSE				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
CCO/1923	Hemangiosarcoma	Solid	B6C3F1	
CE1460 MACA	Mammary Adenocarcinoma	Solid	CE	
CH	Mammary Adenocarcinoma	Solid	Nude C3H	
Cloudman Melanoma	Melanoma	Solid	DBA/1J	See Jax S91
Colon 26	Carcinoma	Solid	BALB/c	
Colon 38	Carcinoma	Solid	C57BL/6	
Crocker Sarcoma				See S180
D ₁ T ₁₀				See LSTRA/DTIC
DBA/2 Spontaneous Tumor M114	Mammary Adenocarcinoma	Solid	DBA/2	
dbrB	Anaplastic Carcinoma	Solid	DBA/1J	See Jax tumors
Egg/Mouse Leukemia	Lymphocytic Leukemia	Ascites	DBA/2, Truslow Egg	
Ehrlich Ascites	Mammary Adenocarcinoma	Solid or Ascites	Various	Several lines
Ehrlich Ascites/6-TG (NSC-752)	Mammary Adenocarcinoma	Ascites	Swiss	Resistant to 6-Thioguanine
Ehrlich Ascites, Tetraploid	Mammary Adenocarcinoma	Ascites	Swiss	
EL-4 Male	Lymphoma	Solid, Spleen C57BL/6 Fragments & Ascites		
E Male Gross	Leukemia	Spleen Homogenate	C57BL/6	
EMT-6	Mammary Adenocarcinoma	Solid	BALB/c	
Ependyoblastoma (Zimmerman)	Ependyoblastoma	Solid	C57BL/6	
FB SAR (A)	Fibrosarcoma	Solid	C3H	
FB SAR (B)	Fibrosarcoma	Solid	C3H	
FCB (C)	Transitional Cell Carcinoma of Bladder	Solid	C57BL/6	

SPECIES: MOUSE				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
Friend Virus Leukemia (FV01)	Reticulum Cell Sarcoma	Solid or Spleen Homogenate	DBA/2	
Furth Tumor				See Carcinoma 1025
Glioma 26	Glioma	Solid	C57BL/6	
Glioma 261	Glioma	Solid	C57BL/6	
Gross Leukemia	Leukemia	Solid	C3H	
Gross Mammary Adenocarcinoma	Mammary Adenocarcinoma	Solid	DBA/2	
Hageman Mastocytoma				See P815
Harding-Passey	Melanoma	Solid	BALB/c	Melanotic and amelanotic types
Hauschka Ascites Tumor	Unknown	Ascites	A/He	
HE10734	Osteogenic Sarcoma	Solid	C3H	
HE10734/FR	Osteogenic Sarcoma	Solid	C3H	
Hepatoma 129 (HE129)	Hepatoma	Solid	C3H or Hybrid	
Hepatoma 134 (HE134, Shear Hepatoma 134)	Hepatoma	Ascites	C3H	
HP	Amelanotic Melanoma	Solid	BALB/cJ	See Jax tumors
H6	Hepatoma	Solid	A/J	See Jax tumors
H2712	Mammary Adenocarcinoma	Solid	C3H/HeJ	See Jax tumors
J-30237	Unknown	Ascites	C3H	
Klein Tumor (TA3)	Mammary Adenocarcinoma	Ascites	A/He or CAF ₁	Several lines
Krebs Ascites Carcinoma	Carcinoma of Inguinal Region	Ascites	C3H or CDBA	
Krebs 2 Carcinoma	Carcinoma of Inguinal Region	Ascites	C3H	
L1210	Lymphoid Leukemia	Ascites or Spleen Homogenate	DBA/2 or CDBA	
L1210/TSC (NSC-729)	Lymphoid Leukemia	Ascites	DBA/2	Resistant to Thio-semicarbazone

SPECIES: MOUSE				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
L1210/MTX (NSC-740)	Lymphoid Leukemia	Ascites or Spleen Homogenate	DBA/2 or CDBA	Treated
L1210/6MP (NSC-755)	Lymphoid Leukemia	Ascites	DBA or Hybrid	Several lines
L1210/L-PAM (NSC-8806)	Lymphoid Leukemia	Ascites	DBA/2 or BDF ₁	Treated
L1210/NSC-19622	Lymphoid Leukemia	Ascites	DBA/2	
L1210/5FU (NSC-19893)	Lymphoid Leukemia	Ascites	BDF ₁	
L1210/CTX (NSC-26271)	Lymphoid Leukemia	Ascites or Spleen Homogenate	DBA/2 or CDBA	
L1210/DF8 (NSC-29630)	Lymphoid Leukemia	Ascites or Spleen Homogenate	DBA/2 or CDBA	Resistant to dichloromethotrexate
L1210/HU (NSC-32065)	Lymphoid Leukemia	Ascites	DBA/2	
L1210/MeGAG (NSC-32946)	Lymphoid Leukemia	Ascites or Spleen Homogenate	CDF ₁	
L1210/NSC-38280	Lymphoid Leukemia	Ascites	CDF ₁	
L1210/DTIC (NSC-45388)	Lymphoid Leukemia	Ascites or Solid	DBA/2 or CDBA	Untreated and treated lines
L1210/TIC (NSC-60339)	Lymphoid Leukemia	Spleen Homogenate	CDBA	Untreated and treated lines
L1210/Ara-C (NSC-63878)	Lymphoid Leukemia	Ascites or Spleen Homogenate	DBA/2 or Hybrid	Untreated and treated lines
L1210/cis-DDP (NSC-119875)	Lymphoid Leukemia	Ascites	DBA/2	Treated
L1210/Anhydro-Ara C (NSC-145668)	Lymphoid Leukemia	Ascites	DBA/2 or Hybrid	Untreated and treated lines
L1210/Ftorafur (NSC-148958)	Lymphoid Leukemia	Ascites	DBA/2 or BDF ₁	Untreated and treated lines
L1210/BCNU (NSC-409962)	Lymphoid Leukemia	Ascites	DBA/2 or Hybrid	Untreated and treated lines
L1210/C95	Lymphoid Leukemia	Ascites or Spleen	CDBA	CTX, MTX, MP resistant

SPECIES: MOUSE				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
		Homogenate		
L1210 FR3 DCM/R 100a	Lymphoid Leukemia	Spleen Homogenate	CDBA	
L1210/FR8/DCM	Lymphoid Leukemia	Spleen Homogenate	CDF ₁	
L1210/FR8 (Folate Reductase)	Lymphoid Leukemia	Spleen Homogenate	CDF ₁	
L1210/M-773	Lymphoid Leukemia	Ascites	DBA/2	Treated
L1210/6MP/6TG	Lymphoid Leukemia	Ascites	CDF ₁	Untreated and treated lines
L1210 variants				See PR ₁ C ₁ T ₅ /NSC-45388, PR ₁ SE ₁ T ₅ and PR ₁ SE ₁ T ₅ /NSC-45388
L4946	Lymphocytic Leukemia	Solid	AKR	
L5178Y	Leukemia	Ascites	DBA/2 or Hybrid	Several lines
L18464	Lymphoma	Solid	C57BL/6	
LAF ₁	Adrenal Cortical Adenocarcinoma	Solid	LAF ₁ /J	
LC-12	Pulmonary Squamous Cell Carcinoma	Solid	BALB/c	
Lewis Lung	Carcinoma	Solid	C57BL	
Lewis Lung/PALA (NSC-224131)	Carcinoma	Solid	C57BL/6	
Lewis Sarcoma T241	Pleiomorphic Cell Sarcoma	Solid	C57BL	
LPC-1	Plasmacytoma	Solid or Ascites	BALB/c	
LS402AX	Teratosarcoma	Solid	C57BL/6 and 129	
LSTRA	Lymphosarcoma	Ascites	BALB/c	Several lines
LSTRA/DTIC (NSC-45388)	Lymphosarcoma	Ascites	BALB/c	Untreated and treated lines
M4898	Lung Adenocarcinoma	Solid	BALB/c	
M5076	Reticulum Cell Sarcoma	Solid or Ascites	C57BL/6	
M5076/L-PAM	Reticulum Cell	Solid	C57BL/6	Treated

SPECIES: MOUSE				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
(NSC-8806)	Sarcoma			
M5076/HMM (NSC-13875)	Reticulum Cell Sarcoma	Solid	C57BL/6	Treated
M5076/cis-DDP (NSC-119875)	Reticulum Cell Sarcoma	Solid	C57BL/6	Treated
M5480	Testicular Carcinoma (Seminoma)	Solid	C57BL/6	
MA13C	Mammary Adenocarcinoma	Solid	C3H	
MA16C	Mammary Adenocarcinoma	Solid	C3H	
MA387	Fusiform Cell Sarcoma	Solid	AKR	
Madison Lung (TA109)	Carcinoma	Solid	BALB/c	
MC-5	Mammary Adenocarcinoma	Spleen	BALB/c	
MC-6 Female	Mammary Adenocarcinoma	Ascites	BALB/c	
MC-11	Mammary Adenocarcinoma	Spleen Homogenate	BALB/c	
MCS-1	Mammary Adenocarcinoma	Solid or Spleen Homogenate	BALB/c	
Mecca (ME61, MLS)	Lymphosarcoma	Solid or Ascites	C3H or AKR	
METH-A	Sarcoma	Ascites	BALB/c	
MLS				See Mecca
Moloney Sarcoma (SV-122-TR4)	Sarcoma	Solid	BALB/c	
MOPC-4	Plasmacytoma	Solid or Ascites	BALB/c	
MOPC-17	Plasmacytoma	Solid	BALB/c	
MOPC-21	Plasmacytoma	Solid or Ascites	BALB/c	
MOPC-28	Plasmacytoma	Solid	BALB/c	
MOPC-30	Plasmacytoma	Solid	BALB/c	
MOPC-31	Plasmacytoma	Solid	BALB/c	
MOPC-41	Plasmacytoma	Solid	BALB/c	

SPECIES: MOUSE				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
MOPC-46	Plasmacytoma	Solid	BALB/c	
MOPC-47	Plasmacytoma	Solid	BALB/c	
MOPC-48	Plasmacytoma	Solid	BALB/c	
MOPC-49	Plasmacytoma	Solid	BALB/c	
MOPC-51	Plasmacytoma	Solid	BALB/c	
MOPC-61	Plasmacytoma	Solid	BALB/c	
MOPC-63	Plasmacytoma	Solid	BALB/c	
MOPC-67	Plasmacytoma	Solid	BALB/c	
MOPC-69	Plasmacytoma	Solid	BALB/c	
MOPC-70	Plasmacytoma	Solid	BALB/c	
MOPC-78	Plasmacytoma	Solid	BALB/c	
MOPC-88	Plasmacytoma	Solid	BALB/c	
MOPC-91	Plasmacytoma	Spleen Homogenate	BALB/c	
MOPC-96	Plasmacytoma	Solid	BALB/c	
MOPC-99	Plasmacytoma	Solid	BALB/c	
MOPC-104	Plasmacytoma	Solid	BALB/c	
MOPC-112	Plasmacytoma	Solid	BALB/c	
MOPC-113	Plasmacytoma	Solid	BALB/c	
MOPC-114	Plasmacytoma	Solid	BALB/c	
MOPC-116	Plasmacytoma	Solid	BALB/c	
MOPC-118	Plasmacytoma	Solid	BALB/c	
MOPC-121	Plasmacytoma	Solid	BALB/c	
MOPC-123	Plasmacytoma	Solid	BALB/c	
MOPC-129	Plasmacytoma	Solid	BALB/c	
MOPC-132	Plasmacytoma	Solid	BALB/c	
MOPC-140	Plasmacytoma	Solid	BALB/c	
MOPC-141	Plasmacytoma	Solid	BALB/c	
MOPC-157	Plasmacytoma	Solid	BALB/c	

SPECIES: MOUSE				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
MOPC-172	Plasmacytoma	Solid	BALB/c	
MOPC-173	Plasmacytoma	Solid	BALB/c	
MOPC-209	Plasmacytoma	Solid	BALB/c	
MPC-1	Plasmacytoma	Solid or Ascites	BALB/c	
MPC-2	Plasmacytoma	Solid or Ascites	BALB/c	
MPC-15	Plasmacytoma	Solid	BALB/c	
MPC-25	Plasmacytoma	Solid	BALB/c	
MPC-26	Plasmacytoma	Solid	BALB/c	
MPC-31	Plasmacytoma	Solid	BALB/c	
MPC-36	Plasmacytoma	Solid	BALB/c	
MPC-37	Plasmacytoma	Solid	BALB/c	
MPC-40	Plasmacytoma	Solid	BALB/c	
MPC-42	Plasmacytoma	Solid	BALB/c	
MPC-44	Plasmacytoma	Solid or Ascites	BALB/c	
MPC-48	Plasmacytoma	Solid	BALB/c	
MPC-49	Plasmacytoma	Solid	BALB/c	
MPC-59	Plasmacytoma	Solid	BALB/c	
MPC-60	Plasmacytoma	Solid	BALB/c	
MPC-63	Plasmacytoma	Solid	BALB/c	
MPC-64	Plasmacytoma	Solid	BALB/c	
MPC-67	Plasmacytoma	Solid	BALB/c	
MPC-73	Plasmacytoma	Solid	BALB/c	
MPC-H	Plasmacytoma	Solid	BALB/c	
MST	Mast Cell	Solid	LAF ₁	
MS-2	Sarcoma	Solid	BALB/c	
MXT	Mammary Ductal Papillary Carcinoma	Solid	BDF ₁	Estrogen Responsive
NK-Lymphoma	Lymphoma	Ascites	C57BL/6	
Nettesheim Lung	Squamous Cell Lung			

SPECIES: MOUSE				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
(LePage Clone KLN205)	Carcinoma	Solid	DBA/2 or Hybrid	
P288	Lymphocytic Leukemia	Solid or Ascites	DBA/2 or CDBA	
P288/MTX (NSC-740)	Lymphocytic Leukemia	Ascites	DBA/2 or BDF ₁	
P388	Lymphocytic Leukemia	Ascites	DBA/2 or CDBA	
P388/MTX (NSC-740)	Lymphocytic Leukemia	Ascites	DBA/2	Treated
P388/ActinomycinD (NSC-3053)	Lymphocytic Leukemia	Ascites	DBA/2	
P388/DON (NSC-7365)	Lymphocytic Leukemia	Ascites	DBA/2	Treated
P388/L-PAM (NSC-8806)	Lymphocytic Leukemia	Ascites	BDF ₁	Treated
P388/5FU (NSC-19893)	Lymphocytic Leukemia	Ascites	BDF ₁	Treated
P388/CPA (NSC-26271)	Lymphocytic Leukemia	Ascites	BDF ₁	Treated
P388/Ara-C (NSC-63878)	Lymphocytic Leukemia	Ascites	BDF ₁	
P388/VCR (NSC-67574)	Lymphocytic Leukemia	Ascites	BDF ₁	Untreated and treated lines
P388/Daunomycin (NSC-82151)	Lymphocytic Leukemia	Ascites	DBA/2	
P388/5-Azacytidine (NSC-102816)	Lymphocytic Leukemia	Ascites	DBA/2	Treated
P388/ADR (NSC-123127)	Lymphocytic Leukemia	Ascites	BDF ₁	Treated
P388/L-Alanosine (NSC-153353)	Lymphocytic Leukemia	Ascites	DBA/2	
P388/Acivicin (NSC-163501)	Lymphocytic Leukemia	Ascites	DBA/2	
P388/Amsacrine (NSC-249992)	Lymphocytic Leukemia	Ascites	DBA/2	
P388/Mitoxantrone (NSC-301739)	Lymphocytic Leukemia	Ascites	DBA/2	
P388/Ara-A + 2'dcF (NSC-404241 + NSC-218321)	Lymphocytic Leukemia	Ascites	BDF ₁	Treated

SPECIES: MOUSE				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
P388/BCNU (NSC-409962)	Lymphocytic Leukemia	Ascites	BDF ₁	Treated
P815 (Hageman Mastocytoma)	Mast Cell Leukemia	Ascites	DBA/2 or Hybrid	
P815/VLB (NSC-49842)	Mast Cell Leukemia	Ascites	DBA/2 or BDF ₁	
P1534	Lymphocytic Leukemia	Spleen Homogenate or Ascites	DBA/2	Several lines
P1798	Lymphosarcoma	Solid or Ascites	BALB/c	
P1798/CR-JS	Lymphoma	Solid	BALB/c	Glucocorticoid resistant, treated
P1798/CS-JS	Lymphoma	Solid	BALB/c	Glucocorticoid sensitive
PAN 02	Pancreas	Solid	C57BL/6	
PAN 03	Pancreas	Solid	C57BL/6	
PR ₁ C ₁ T5/NSC-45388	Lymphoid Leukemia	Ascites	CDF ₁	L1210 variant; treated
PR ₁ SE ₁ T5	Lymphoid Leukemia	Ascites	CDF ₁	L1210 variant
PR ₁ SE ₁ T5/NSC-45388	Lymphoid Leukemia	Ascites Homo- genate	CDF ₁	L1210 variant; treated
R-26	Unknown	Ascites	CDF ₁	
R-46	Unknown	Ascites	CDF ₁	
R-53	Unknown	Ascites	CDF ₁	
R-74	Unknown	Ascites	CDF ₁	
RBL-5 (Rauscher Virus Induced Transplantable Tumor-5)	Leukemia	Ascites	C57BL/6	
RC-2	Renal Adenocarcinoma	Ascites	CDF ₁	
Reif-Allen Tumor	Thymoma	Ascites	AKR	
RPC-5	Plasmacytoma	Solid	BALB/c	
RPC-9	Plasmacytoma	Solid or Ascites	BALB/c	
RPC-20	Plasmacytoma	Solid or Ascites	BALB/c	
S37	Pleomorphic Cell Sarcoma	Ascites	BALB/c, Nonspecific	See Jax tumors

SPECIES: MOUSE				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
S180 (Crocker, S III)	Pleomorphic Cell Sarcoma	Solid or Ascites	BALB/c, Swiss or hybrids	See Jax tumors
Sa 1	Spindle Cell Sarcoma	Solid	A/J	See Jax tumors
Sa D2	Fibrosarcoma	Solid	DBA/2J	See Jax tumors
Shear Hepatoma 134				See Hepatoma 134
SJL/JW	Reticulum Cell Sarcoma	Spleen Homogenate	SJL/JW	
Spontaneous Adrenal	Adrenal	Solid	CE/J	
Spontaneous DBA/2 Mammary	Mammary Adenocarcinoma	Solid	DBA/2	
Spontaneous Mammary	Mammary Adenocarcinoma	Solid	DBA/2	
SV-122-TR4				See Moloney Sarcoma
TA3				See Klein tumor
TA109				See Madison Lung
T1699	Mammary Adenocarcinoma	Solid	DBA/2J	See Jax tumors
T1703	Mammary Adenocarcinoma	Solid	DBA/1J	See Jax tumors
X5563	Unknown	Solid	C3H/He	
YPC-1	Plasmacytoma	Ascites	BALB/c	
Zimmerman Ependymoblastoma				See Ependymoblastoma
6C3HED (Gardner)	Lymphosarcoma	Ascites	C3H	Several lines. See Jax tumors
6C3HED/AR Res.	Lymphosarcoma	Spleen Homogenate	C3H	
102A	Unknown	Ascites	CDF ₁	
1247	Mammary Adenocarcinoma	Solid	BALB/c	
36230 TLT	Hemangio-endothelioma	Solid	C57BL/6J	
36257 TTT	Fibrosarcoma	Solid	BALB/cAnN	
38290 TTT	Fibrosarcoma	Solid	BALB/cAnN	

SPECIES: MOUSE				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
42021 TCT	Hemangio-endothelioma	Solid	BALB/cAnN	
42022 TST	Hemangio-endothelioma	Solid	BALB/cAnN	
42052 TST	Hemangio-endothelioma	Solid	C57BL/6J	
42076 TST	Hemangio-endothelioma	Solid	C57BL/6J	
44316 TST	Hemangio-endothelioma	Solid	BALB/cAnN	
44316 LTST	Hemangio-endothelioma	Solid	BALB/cAnN	
44347 TST	Hemangio-endothelioma	Solid	BALB/cAnN	
46362 TTT	Fibrosarcoma	Solid	C57BL/6J	
46363 TTT	Fibrosarcoma	Solid	BALB/cAnN	
70429	Plasmacytoma	Ascites	C3H	
70429/Azaserine (NSC-3425)	Plasmacytoma	Ascites	C3HF/LW	
91632	Reticulum Cell Sarcoma	Solid	C57BL/Kaplan	

MOUSE TUMORS FROM THE JACKSON LABORATORY

MOUSE TUMORS FROM THE JACKSON LABORATORY IN CRYOPRESERVATION IN THE DCTD TUMOR REPOSITORY

Tumor Designation	Tumor Type	Host Strain	Transplantation Frequency (days)	Host Survival (days)	Lag Time* (days)	Strain of Origin	Sex of Origin	MRI Bank #	MAP Test ⁺
dbrB	Anaplastic Carcinoma	DBA/1J	7	7-9	5-7	DBA		J-730	LDH+
SaD2	Fibrosarcoma	DBA/2J	10	19-12	8-15	DBA/2J		J-765	LDH+
H6	Hepatoma	A/J	10-14	14-44	7-9	A/J		J-750	LDH+
BW7756	Hepatoma	C57L/J	14	13-31	12-16	C57L/J		J-748	LDH+
6C3HED (GL-1)	Lymphosarcoma	C3H/HeJ	7	7-9	16	C3h	--	J-744	LDH+
BW10232	Mammary Adenocarcinoma	C57BL/6J	10	23-30	11	C57BL/6J		J-762	LDH+
C3HBA	Mammary Adenocarcinoma	C3H/HeJ	10	39-77	11-16	C3H/An		H-758	LDH+
H2712	Mammary Adenocarcinoma	C3H/HeJ	7	14-27	11-19	C3H/HeHu		J-731	LDH+
CaD1	Mammary Adenocarcinoma	DBA/1J	10	25-43	9-17	DBA/1J		J-742	LDH+
T1703	Mammary Adenocarcinoma	DBA/1J	10	47-74	9-12	DBA/1Hu		J-737	LDH+
T1699	Mammary Adenocarcinoma	DBA/2J	10	19-39	8-10	DBA/2J		J-736	LDH+
HP	Melanoma (amelanotic)	BALB/cJ	14	28-77	19-23	nonlabred	--	J-746	LDH+
B16	Melanoma (amelanotic)	C57BL/6J	10	24-44	15-21	C57BL/6J	--	J-753	LDH+
S91	Melanoma (melanotic)	DBA/1J	17-21	49-98	16-18	DBA (Snell)	--	J-749	LDH+
C1498	Myeloid Leukemia	C57BL/6J	7	9-16	7-9	C57BL	--	J-738	LDH+
BW8883	Pituitary	C57L/J	60	182-273	65-73	C57L/J	--	J-756	LDH+

Tumor Designation	Tumor Type	Host Strain	Transplantation Frequency (days)	Host Survival (days)	Lag Time* (days)	Strain of Origin	Sex of Origin	MRI Bank #	MAP Test [†]
BW8685	Pituitary	C57BR/dcJ	90-120	210-238	395	C57BR/cdj		J-794	LDH+
S180	Pleomorphic Sarcoma	BALB/cJ	10	21-31	9-11	"white" mouse		J-757	LDH+
S37	Pleomorphic Sarcoma	DBA/1J	7	21-28	6-13	"stock" mouse		J-759	LDH+
C1300	Round cell (Neuroblastoma?)	A/J	10	19-32	14-21	A albino	--	J-734	LDH+ MHV+
Sal	Spindle-cell Sarcoma	A/J	7	9-15	7-9	A albino	--	J-733	LDH+

*Length of lag phase before measurable tumor growth (5 mm average diameter) is evident in the first passage post thaw.

[†]MAP Test - Murine Antigen Profile for 12 common viruses: PVH, Rco 3, Sendai, GDVII, K, Polyoma, MVH, MAB, MHV, LCM, Ectromelia, LDH. Only positive results are listed.

Adapted from Jax Notes, No. 424, December 1975.

RABBIT TUMORS

SPECIES: RABBIT				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
Brown-Pearce	Carcinoma (Epithelioma)	Solid	New Zealand White or Dutch	
VX-2 (V2)	Skin Carcinoma	Solid	New Zealand White or Dutch	

RAT TUMORS

SPECIES: RAT				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
AA Ascites	Spontaneous Ascites	Ascites	Wistar	
ATC 64	Thyroid Carcinoma	Solid	Fischer 344	
9A Ascites	Spontaneous Ascites	Ascites	Inbred PA	
A546 (DMBZ Attenuated)	Unknown	Solid	Fischer 344	
A920 (Tetramin Attenuated Resistant)	Unknown	Solid	Fischer 344	
A1011	Unknown	Solid	Fischer 344	
A1131-AR	Unknown	Solid	Fischer 344	
A1138-AL	Unknown	Solid	Fischer 344	
A1140-CL-10	Unknown	Solid	Fischer 344	
BT/M520	Fibrosarcoma	Solid	Marshall 520	
Carcinosarcoma	Carcinosarcoma (skin) (18-day embryos)	Solid	Sprague-Dawley Derived Holtzman	
CA 20948	Pancreatic Acinar Carcinoma	Solid	Wistar	
CCO 1865	Mesothelioma	Solid	Fischer 344	
CSE	Fibrosarcoma	Solid	Fischer 344	H-1
DD81-23	T cell lymphoma	Mince	Fischer 344	
DL Wells-17	T cell lymphoma	Mince	Fischer 344	
DMBA	Mammary Adenocarcinoma	Solid	Fischer 344	Several lines
DSL62-38	Pancreatic Tumor	Solid	Wistar Lewis	
Dunning Leukemia	Atypical Monocytic Leukemia	Solid or Ascites	Fischer 344	H-1 H-1, KRV
Dunning Leukemia/ NSC-755 (6-MP)	Atypical Monocytic Leukemia	Solid	Fischer 344	
Dunning Leukemia/ NSC-3088 (chlor-ambucil)	Atypical Monocytic Leukemia	Solid	Fischer 344	
Dunning Leukemia/ NSC-10107 (nitromin)	Atypical Monocytic Leukemia	Solid	Fischer 344	
Dunning Leukemia/	Atypical Monocytic	Solid	Fischer 344	

SPECIES: RAT				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
NSC-13875 (HMM)	Leukemia			
Dunning Leukemia/ NSC-17261 (benzo- quinone)	Atypical Monocytic Leukemia	Solid	Fischer 344	
Dunning Leukemia/ NSC-23892 (dimethyl- benzimidazole)	Atypical Monocytic Leukemia	Solid	Fischer 344	
Dunning Leukemia/ NSC-26980 (mitomycin C)	Atypical Monocytic Leukemia	Solid	Fischer 344	
Dunning Leukemia/ NSC-29422 (thio- guanosine)	Atypical Monocytic Leukemia	Solid	Fischer 344	
Dunning Leukemia/ NSC-45059 (o-acetyl- tetramin)	Atypical Monocytic Leukemia	Solid	Fischer 344	
Dunning Leukemia/ NSC-51845 (cyclo- hexylamine)	Atypical Monocytic Leukemia	Solid	Fischer 344	
Flexner-Jobling	Seminal Vesicle Adenocarcinoma	Solid	Nonspecific, Sprague- Dawley or Wistar	
Fran Tumor	Ovarian Carcinoma	Ascites	Sprague-Dawley	
GBT/W	Glial Tumor	Solid	Wistar	
HB Lynch-Fibroma 522	Fibroma	Solid	Fischer 344	
Hepatoma NK				See Novikoff Hepatoma
HMC	Histiocytoma	Solid	Fischer 344	
H-372	Leydig	Solid	Fischer 344	
H-540	Leydig	Solid	Fischer 344	
Iglesias	Ovarian Carcinoma	Solid	ACI	
IRS 9802	Spindle Cell Sarcoma	Solid	Fischer 344	
LC-18	Hepatoma	Solid	Fischer 344	
Lymphoma 8	Lymphoma	Solid, Spleen Homogenate, Whole Blood	Lewis	
L.T.W. (Furth)	Leydig	Solid	Wistar	
MAMF2-TC	Fibrosarcoma	Solid	Fischer 344	

SPECIES: RAT				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
MET 149-2	Adenocarcinoma	Solid	Fischer 344	
Moore Sarcoma #1	Sarcoma	Solid	Wistar	
Morris Hepatomas	Hepatoma	Solid	Buffalo	
16	Hepatoma	Solid	Buffalo	
20	Hepatoma	Solid	Buffalo	
44	Hepatoma	Solid	Buffalo	
3924A	Hepatoma	Solid	ACI	
5123	Hepatoma	Solid	Buffalo	
7777	Hepatoma	Solid	Buffalo	
7800	Hepatoma	Solid	Buffalo	
8999	Hepatoma	Solid	Buffalo	
9618A	Hepatoma	Solid	Buffalo	
MNU-Buffalo	Mammary Carcinoma	Solid	Buffalo	Several lines
MtT	Anterior Pituitary	Solid	Fischer, Wistar	Several lines
MT/W9	Mammary Adenocarcinoma	Solid	Wistar	Several lines
MT/W449	Mammary Adenocarcinoma	Solid	Wistar	Several lines
Murphy-Sturm Lymphosarcoma (MSL)	Lymphosarcoma	Solid	CRL, Wistar, Fischer 344, Sprague-Dawley	
NBW-37	T cell lymphoma	Mince	Fischer 344	
Novikoff Hepatoma (Hepatoma NK)	Hepatoma	Solid or Ascites	Random bred albino (Sprague-Dawley weanlings)	
NS104	Rhabdomyosarcoma	Solid	Fischer 344	
OR-16-3	Thymus Tumor	Solid	Fischer 344	
R35	Mammary Adenocarcinoma	Solid	Holtzman	
R3259	Giant Cell Sarcoma	Solid	Fischer 344	
Rice D6	Leydig	Solid	Fischer 344	
Rice 500	Leydig	Solid	Fischer 344	

SPECIES: RAT				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
Riejoel	Thyroid Adenocarcinoma	Solid	Fischer 344	
RNC 259	Pheochromocytoma	Solid	NEDH	
RNC 288	Insulinoma	Solid	NEDH	
RNK-16	LGL Leukemia	Solid or Spleen Homogenate	Fischer 344	
R3149	Leukemia	Solid	Fischer 344	
R3230AC	Mammary Adenocarcinoma	Solid	Fischer 344	
R3327	Prostate	Solid	Copenhagen 2331	
R3327 (Pap)	Prostate	Solid	Copenhagen 2331	
3M2N	Mammary Squamous Cell Carcinoma	Solid	Fischer 344	
Shay Leukemia	Myelogenous Leukemia	Solid	Sprague-Dawley	
SMT-2A	Mammary Carcinoma	Solid	Fischer 344	
Swarm	Chondrosarcoma	Solid	Sprague-Dawley	
TR.CLXXXVIII	Melanoma	Solid	ACI	
TR.DCCXLIII	Pituitary	Solid	ACI	
Walker 256	Carcinosarcoma	Solid or Ascites	Sprague-Dawley	
Yoshida Hepatoma	Hepatoma	Ascites	Sprague-Dawley	
Yoshida Sarcoma	Sarcoma	Solid or Ascites	Holtzman, S-D	
68-2	Alveolar/Bronchiolar Carcinoma	Solid	Fischer 344	
13762	Mammary	Solid or Ascites	Fischer 344	Several lines
13762-FS	Fibrosarcoma	Solid	Fischer 344	
23 Methapyrilene	Hepatocellular Carcinoma	Solid	Fischer 344	
29 Methapyrilene	Hepatocellular Carcinoma	Solid	Fischer 344	
33 Methapyrilene	Hepatocellular Carcinoma	Solid	Fischer 344	
2982	Olfactory	Solid	Fischer 344	

SPECIES: RAT				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
	Neuroblastoma			
11095	Prostate	Solid	Fischer 344	

RAT TUMORS FROM DR. ROBERT NOBLE (ENDOCRINE-RESPONSIVE)

SPECIES: RAT (tumors received from Dr. Robert Noble)

Information concerning the endocrinologic characteristics of the various tumor systems indicated in the "comments" is that supplied by Dr. Noble.

SPECIES: RAT (tumors received from Dr. Robert Noble)				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
1 Cvx-34A(1)	Cervical Carcinoma	Solid	NB	
1 Cvx-44Z	Cervical Carcinoma	Solid	NB	Estrogen dependent
1 Lym-206	Lymphoma	Solid	NB	Hormone stimulated
1 Lym-209(A)	Lymphoma	Solid	NB	Hormone stimulated, VLB sensitive
1 Lym-214	Lymphosarcoma	Solid	NB	Hormone stimulated
1 Og-3	Osteogenic Sarcoma	Solid	NB	
1 Pan-14Ax(1)	Adenocarcinoma	Solid	NB	Hormone stimulated
1 Sal-23(2)	Salivary Gland, Secretory Tumor	Solid	NB	Estrogen dependent
1 Tes-13E	Leydig Cell Carcinoma	Solid	NB	Estrogen dependent
1 TES-15E	Leydig Cell Carcinoma	Solid	NB	Estrogen dependent
Kid-27B	Kidney Adenocarcinoma	Solid	NB	
2 Lym-11(a)	Metaplastic, Adeno- carcinoma Fibroblast Overgrowth	Solid	NB	Estrogen dependent
2-Pan-6A	Pituitary Adenoma	Solid	NB	Hormone stimulated
2-Pr-7A	Prostatic Adenocarcinoma	Solid	NB	Estrogen dependent
2 Pr-9F	Prostatic Adenocarcinoma	Solid	NB	Estrogen dependent
2 Pr-12	Prostatic Adenocarcinoma	Solid	NB	Estrogen dependent
2 Pr-112Bx(1)	Prostatic Carcinoma, Scirrhou	Solid	NB	
2 Pr-114B	Prostatic Adenocarcinoma	Solid	NB	Estrogen dependent
2 Pr-121D(1)	Prostatic Carcinoma, Secretory	Solid	NB	Androgen dependent
2 Pr-121D(1)/R	Prostatic Carcinoma	Solid	NB	Resistant to testosterone
2 Sk-103	Melanoma	Solid	NB	
2 Ut-10(5)	Fibroma	Solid	NB	Estrogen dependent
3 Kid-13	Kidney Adenocarcinoma	Solid	NB	
3 Lym-19	Lymphosarcoma	Solid	NB	
4 Pan-6	Adenocarcinoma	Solid	NB	

SPECIES: RAT (tumors received from Dr. Robert Noble)				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
4 Sk-3A(3)Z	Squamous Cell Carcinoma	Solid	NB	
4 Ut	Hemangiosarcoma	Solid	NB	
4 Ut-6(2)	Fibrosarcoma	Solid	NB	Estrogen dependent or hormone stimulated
5 Pan-7	Undifferentiated Pancreatic Carcinoma	Solid	NB	
5 Sal	Undifferentiated Carcinoma	Solid	NB	
5 Sk-3	Melanoma	Solid	NB	
5 Ut-2	Uterine Adenocarcinoma	Solid	NB	Estrogen dependent (?)
6 Pan-4	Undifferentiated Pancreatic Carcinoma	Solid	NB	
7 Ut-13	Endometrial Adenocarcinoma	Solid	NB	
8 Lym-9(1)	Lymphosarcoma	Solid	NB	VLB resistant
8 Lym-108(1)	Lymphatic Leukemia	Solid	NB	VLB resistant
9 Lym-23	Lymphosarcoma	Solid	NB	
10 Lym-4	Negative Spleen	Solid	NB	
11 Lym-9	Lymphosarcoma	Solid	NB	
13 Pr-5	Prostatic Carcinoma, Undifferentiated	Solid	NB	Estrogen pellet implant required
14 Lym-5	Lymphosarcoma stimulated	Solid	NB	Hormone
14 Pr-5	Prostatic Carcinoma	Solid	NB	
15 Pr-2	Prostatic Adenocarcinoma	Solid	NB	
16 Pr-3	Prostatic Adenocarcinoma	Solid	NB	
17 Lym-4	Lymphosarcoma implant required	Solid	NB	Estrogen pellet
17 Lym-5	Leukemia	Solid	NB	
18 Lym-6	Lymphosarcoma	Solid	NB	Estrogen dependent
19 Lym-3	Lymphosarcoma	Solid	NB	Estrogen dependent
19 Pr-19	Prostatic Fibroadenoma	Solid	NB	
20 Pr-1	Prostatic Fibroadenoma	Solid	NB	
20 Lym-3	Lymphosarcoma	Solid	NB	Estrogen pellet implant required

SPECIES: RAT (tumors received from Dr. Robert Noble)				
Tumor Designation	Histologic Type	Form	Strain of Origin/ Transplant	Comments
21 Pr-9	Prostatic Carcinoma	Solid	NB	
22 Pr-8	Prostatic Adenocarcinoma	Solid	NB	
23 Pr-7	Prostatic Carcinoma	Solid	NB	
24 Pr-2	Prostatic Carcinoma	Solid	NB	
25 Pr-3	Prostatic Adenocarcinoma	Solid	NB	

***IN VITRO* ESTABLISHED CELL LINES**

HUMAN IN VITRO CELL LINES				
Designation	Tissue of Origin	Histologic Type	Growth Medium	Remarks
ACHN	Kidney	Renal Cell Carcinoma	RPMI 1640	From Dr. Schmid
A2780	Ovary	Adenocarcinoma	RPMI 1640	From Dr. Hamilton
A498	Kidney	Renal Cell Carcinoma	RPMI 1640	ATCC
A549	Lung	Non-small Cell	RPMI 1640	ATCC
A704	Kidney	Renal Cell Carcinoma	RPMI 1640	ATCC
BT-549	Breast	Adenocarcinoma	RPMI 1640	ATCC
CAKI-1	Kidney	Renal Cell Carcinoma	RPMI 1640	From Dr. Loveless
CCRF-CEM	Lymph	Leukemia	RPMI 1640	ATCC
CCRF-SB	Lymph	Leukemia	RPMI 1640	ATCC
CHA-59	Bone	Osteosarcoma	RPMI 1640	From Drs. Shoemaker and McLachlan
COLO 205	Colon	Adenocarcinoma	RPMI 1640	ATCC
DMS-114	Lung	Small Cell	RPMI 1640	From Dr. Pettengill
DU-145	Prostate	Carcinoma	RPMI 1640	ATCC
EKVX	Lung	Adenocarcinoma	RPMI 1640	From Dr. Fodstad
HCC-2998	Colon	Adenocarcinoma	RPMI 1640	From Dr. Fidler
HCT-15	Colon	Carcinoma	RPMI 1640	ATCC
HCT-116	Colon	Adenocarcinoma	RPMI 1640	ATCC
HELA	Cervix	Carcinoma		
HOP-18	Lung	Large Cell Carcinoma	RPMI 1640	From Drs. Liu/Casero
HOP-62	Lung	Adenocarcinoma	RPMI 1640	From Drs. Liu/Casero
HOP-92	Lung	Large Cell	RPMI 1640	From Dr. Liu
HL-3		Lymphoma		
HL-4		Lymphoma		
HL-60	Ascites	Pro-myelocytic Leukemia	RPMI 1640	From E. Jensen
H-MESO-1		Mesothelioma	RPMI 1640	
HS 578T	Breast	Adenocarcinoma	RPMI 1640	ATCC
HS 913T	Lung	Mixed Cell	RPMI 1640	ATCC
HT-29	Colon	Adenocarcinoma	RPMI 1640	ATCC
IGR-OV1	Ovary	Adenocarcinoma	RPMI 1640	From Dr. Benard

HUMAN IN VITRO CELL LINES

Designation	Tissue of Origin	Histologic Type	Growth Medium	Remarks
KB-ADL #12	Oral Cavity	Epidermoid	EMEM	
KM-12	Colon	Adenocarcinoma	RPMI 1640	From Dr. Fidler
KM 20L2	Colon	Adenocarcinoma	RPMI 1640	From Dr. Fidler
K-562	Lymph	Leukemia	RPMI 1640	ATCC
LOVO	Colon	Adenocarcinoma	RPMI 1640	ATCC
LOX IMVI	Lymph Node Metastasis	Amelanotic Melanoma	RPMI 1640	From Dr. Fodstad
LXFL 529**	Lung	Large Cell Carcinoma	RPMI 1640	From Dr. Fiebig**
MALME-3M	Lung Metastasis	Melanoma	RPMI 1640	ATCC
MCF7	Breast	Adenocarcinoma	RPMI 1640	From Dr. Cowan
MDA-MB-231	Breast	Adenocarcinoma	RPMI 1640	From Dr. Moore
MDA-MB-435	Melanoma	Adenocarcinoma	RPMI 1640	From Dr. Steeg
MDA-MB-468	Breast	Adenocarcinoma		
MOLT-4	Lymph	Leukemia	RPMI 1640	ATCC
MX-1	Breast	Carcinoma	RPMI 1640	From Dr. Giovannelli
M14		Amelanotic Melanoma	RPMI 1640	From Dr. Kern
M19-MEL		Amelanotic Melanoma	RPMI 1640	From Dr. Kern
NC-37	Lymphoblast	Normal		
NCI-H23	Lung	Adenocarcinoma	RPMI 1640	From Dr. Gazdar
NCI-H69	Lung	Small Cell Carcinoma	RPMI 1640	From Dr. Gazdar
NCI-H82	Lung	Small Cell Carcinoma	RPMI 1640	From Dr. Gazdar
NCI-H125	Lung	Adenosquamous Carcinoma	RPMI 1640	From Dr. Gazdar
NCI-H226	Lung	Squamous Cell	RPMI 1640	From Dr. Gazdar
NCI-H292	Lung	Adenosquamous Carcinoma	RPMI 1640	From Dr. Gazdar
NCI-H322M	Lung	Adenocarcinoma	RPMI 1640	From Dr. Gazdar
NCI-H358M	Lung	Bronchioalveolar Carcinoma	RPMI 1640	From Dr. Gazdar
NCI-H460	Lung	Large Cell	RPMI 1640	From Dr. Gazdar
NCI-H522	Lung	Adenocarcinoma	RPMI 1640	From Dr. Gazdar
NCI/ADR-RES	Ovary	Adenocarcinoma	RPMI 1640	From Dr. Cowan Please see JNCI correspondence on page 64
OV	Ovary			

HUMAN IN VITRO CELL LINES				
Designation	Tissue of Origin	Histologic Type	Growth Medium	Remarks
OVCAR-3	Ovary	Adenocarcinoma	RPMI 1640	From Drs. Ozols and Hamilton
OVCAR-4	Ovary	Adenocarcinoma	RPMI 1640	From Drs. Ozols and Hamilton
OVCAR-5	Ovary	Adenocarcinoma	RPMI 1640	From Drs. Ozols and Hamilton
OVCAR-8	Ovary	Adenocarcinoma	RPMI 1640	From Drs. Ozols and Hamilton
PC-3	Prostate	Carcinoma	RPMI 1640	From Dr. Kaighn
PC-3/M	Prostate	Carcinoma	RPMI 1640	From Dr. Kaighn
RPMI-7951		Melanoma	RPMI 1640	ATCC
RPMI-8226	Lymph	Leukemia	RPMI 1640	ATCC
RXF 393	Kidney	Renal Cell Carcinoma	RPMI 1640	From Dr. Fiebig
RXF 631	Kidney	Renal Cell Carcinoma	RPMI 1640	From Dr. Fiebig
SF-268	CNS	Glioblastoma	RPMI 1640	From Dr. Rosenblum
SF-295	CNS	Glioblastoma	RPMI 1640	From Dr. Rosenblum
SF-539	CNS	Glioblastoma	RPMI 1640	From Dr. Rosenblum
SHP-77	Lung	Small Cell Carcinoma	RPMI 1640	From Dr. Fisher
SK-OV-3	Ovary	Adenocarcinoma	RPMI 1640	ATCC
SK-MEL-2		Melanoma	RPMI 1640	ATCC
SK-MEL-5		Melanoma	RPMI 1640	ATCC
SK-MEL-28		Melanoma	RPMI 1640	ATCC
SK-MES-1	Lung	Squamous Cell Carcinoma	RPMI 1640	ATCC
SN12A1	Kidney	Renal Cell Carcinoma	RPMI 1640	From Dr. Fidler
SN12C	Kidney	Renal Cell Carcinoma	RPMI 1640	From Dr. Fidler
SN12K1	Kidney	Renal Cell Carcinoma	RPMI 1640	From Dr. Fidler
SN12L1	Kidney	Renal Cell Carcinoma	RPMI 1640	From Dr. Fidler
SN12S1	Kidney	Renal Cell Carcinoma	RPMI 1640	From Dr. Fidler
SNB-7	CNS	Glioblastoma	RPMI 1640	From Dr. Kornblith
SNB-19	CNS	Glioblastoma- (Same as U251)	RPMI 1640	From Dr. Kornblith
SNB-75	CNS	Glioblastoma	RPMI 1640	From Dr. Kornblith
SNB-78	CNS	Astrocytoma	RPMI 1640	From Dr. Kornblith
SR	Pleural effusion	Lymphoma	RPMI 1640	From Dr. Urba

HUMAN IN VITRO CELL LINES				
Designation	Tissue of Origin	Histologic Type	Growth Medium	Remarks
SW-620	Colon		RPMI 1640	ATCC
T-47D	Breast		RPMI 1640	Not distributed to commercial firms or for commercial purposes
TK-10	Kidney	Renal Cell Carcinoma	RPMI 1640	From Dr. Clayman
UACC-62		Melanoma	RPMI 1640	From Dr. Leibowitz
UACC-257		Melanoma	RPMI 1640	From Dr. Leibowitz
UCSD 242L		Melanoma	RPMI 1640	From Dr. Taetle
UCSD 354L		Melanoma	RPMI 1640	From Dr. Taetle
UO-31	Kidney	Renal Cell Carcinoma	RPMI 1640	From Dr. Linehan
U-251	CNS	Glioblastoma- (Same as SNB-19)	RPMI 1640	From Dr. Bigner
WIDR	Colon	Adenocarcinoma	RPMI 1640	ATCC
XF 498	CNS	Glioblastoma	RPMI 1640	From Dr. Fiebig
786-0	Kidney	Renal Cell Carcinoma	RPMI 1640	From Dr. Williams

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NCI ANTI-CANCER CELL LINE PANEL						
Cell Line	Sex	Age	Histology	Comment	Treatment	Source
COLON						
COLO 205	M	70	Adenocarcinoma	Can Res 38: 1345-1355, 1978		
HCC-2998			Carcinoma		N	
HCT-15			Adenocarcinoma	Can Res 39: 1020-1025, 1979		
HCT-116			Carcinoma	Can Res 41: 1761-1756, 1981		
HT29	F	44	Adenocarcinoma, GR III	Human Tumor Cells In Vitro: 115-159, 1975		Primary
KM12			Adenocarcinoma	Can Res 48: 1943-1948, 1988	N	
SW-620	M	51	Adenocarcinoma	Can Res 36: 4562-4569, 1976		Metastasis
CNS						
SF-268	F	24	Anaplastic Astrocytoma	Acta Neuropathol 75: 92-103, 1987		
SF-295	F	67	Glioblastoma-Multiforme	Acta Neuropathol 75: 92-103, 1987		
SF-539				J Neuropathol Exp Neurol 40: 201- 229, 1981		
SNB-19	M	47	Glioblastoma (same as U251)	Cancer 47: 255, 1981	N	
SNB-75	F		Astrocytoma		N	
U251	M	75	Glioblastoma (same as SNB-19)	J Neuropathol Exp Neurol 40: 410-427, 1981		
LEUKEMIA						
CCRF-CEM	M	4	Acute Lymphoblastic Leukemia	Can Res 18: 522-529, 1965		
HL-60(TB)	F	36	Promyelocytic Leukemia	Nature 270: 347-349, 1977		PBL
K-562	F	53	Chronic Myelogenous Leukemia	Blood 45: 321-334, 1975		Pleural Effusion
MOLT-4	M	19	Acute Lymphoblastic Leukemia	JNCI 49: 891-895, 1972		PB
RPMI-8226	M	61	Myeloma	Proc Soc Exp Biol Med 125: 1246-1250, 1967		PB
SR	M	11	Large Cell, Immunoblastic		Y	
LUNG						
A549/ATCC	M	58	Adenocarcinoma	JNCI 51: 1417-1423, 1973		Primary
EKVX	M		Adenocarcinoma			
HOP-62	F	60	Adenocarcinoma		N	
HOP-92	M	62	Large Cell, Undifferentiated		N	
NCI-H23			Adenocarcinoma	Can Res 45: 2913-2923, 1985	N	
NCI-H226			Squamous	Can Res 45: 2913-2923, 1985		

NCI ANTI-CANCER CELL LINE PANEL						
Cell Line	Sex	Age	Histology	Comment	Treatment	Source
LUNG (continued)						
NCI-H322M			Small Cell Bronchioalveolar Carcinoma		N	
NCI-H460	M		Large Cell Carcinoma	Science 246: 491-494, 1989	N	Pleural Effusion
NCI-H522			Adenocarcinoma	Can Res 45: 2913-2923, 1985		
MAMMARY						
MCF7	F	69	Adenocarcinoma	JNCI 51: 1409-1417, 1973	Y	
HS 578T	F	74	Carcinosarcoma	JNCI 58: 1795-1806, 1977		Primary
MDA-MB-231	F	51	Adenocarcinoma	JNCI 53: 661-674, 1974	Y	
MDA-MB-468	F	51	Adenocarcinoma	Cancer Res 40: 3118-3129, 1980		
BT-549	F	72	Papillary Infiltrating Ductal Carcinoma	No Publication		Metastasis
T-47D*	F	54	Infiltrating Ductal Carcinoma	Eur J Cancer 15: 659-670, 1979		*not for commercial use
MELANOMA						
LOX IMVI			Malignant Amelanotic Melanoma	Int J Cancer 41: 442-449, 1988		
M14						
MALME-3M	M	43	Malignant Melanoma	Human Tumor Cells In Vitro, pp. 115-159, 1975		Metastasis
MDA-MB-435	F	31	Adenocarcinoma	Can Res 40: 3118-3129, 1980	N	
SK-MEL-2	M	60	Malignant Melanoma	Human Tumor Cells In Vitro, pp. 115-159, 1975		Metastasis
SK-MEL-5			Malignant Melanoma	PNAS 73: 3278-3282, 1976		Metastasis
SK-MEL-28			Malignant Melanoma	PNAS 73: 3278-3282, 1976		
UACC-62						
UACC-257						
OVARIAN						
IGR-OV1	F	47	Cystoadenocarcinoma	Can Res 45: 4970-4979, 1985	N	
NCI/ADR-RES	F		Adenocarcinoma	JNCI 90(11): 6/3/1998		
OVCAR-3	F	60	Adenocarcinoma	Can Res 43: 5379-5389, 1983	Y	Ascites
OVCAR-4	F	42	Adenocarcinoma	Sem Oncol 11: 285-298, 1984	Y	
OVCAR-5	F	67	Adenocarcinoma	Sem Oncol 11: 285-298, 1984	N	
SK-OV-3	F	64	Adenocarcinoma	Human Tumor Cells In Vitro, pp. 115-159,	Y	Ascites

NCI ANTI-CANCER CELL LINE PANEL						
Cell Line	Sex	Age	Histology	Comment	Treatment	Source
				1975		
PROSTATE						
DU-145	M	69	Carcinoma	Int J Cancer 21: 274-281, 1978	Y	
PC-3	M	62	Adenocarcinoma	Invest Urol 17: 16-23, 1979	Y	Metastasis
RENAL						
786-O	M	58	Adenocarcinoma	In Vitro 12: 623-627, 1976	N	
A498	F	52	Adenocarcinoma	JNCI 51: 1417-1423, 1973		
ACHN	M	22	Renal Cell Carcinoma	Can Res 42: 4948-4953, 1982	Y	Pleural Effusion
CAKI-1	M	49	Clear Cell Carcinoma	Human Tumor Cells In Vitro, pp. 115-159, 1975	Y	Metastasis
RXF 393	M	54	Poorly Differentiated Hypernephroma	Contrib Oncol 42, 1992	N	
SN12C	M	43	Carcinoma	Can Res 46: 4109-4115, 1986		
TK-10	M	43	Spindle Cell Carcinoma	Can Res 47: 3856-3862, 1987	N	
UO-31			Carcinoma		N	

CORRESPONDENCE

Cell Line Designation Change: Multidrug-Resistant Cell Line in the NCI Anticancer Screen

Since 1990, the Developmental Therapeutics Program (DTP) of the National Cancer Institute (NCI) has screened over 60,000 compounds and a larger number of natural product extracts for their capacity to inhibit the growth of 60 different human tumor cell lines (1). These cell lines have been maintained in cryopreservation and in culture, and they have been subjected to strict quality controls, including adventitious agent testing, human isoenzyme analysis, karyology, morphological and immunocytochemical characterization (2), and DNA fingerprinting. One of these cell lines, previously designated as MCF-7/ADR-RES, has been included in the *in vitro* cell line screening panel because of its stable multidrug-resistant (MDR) phenotype (3) characterized by high levels of MDR-1 and P-glycoprotein expression (4,5). Recently, we submitted cell lines from the screening panel for DNA fingerprinting analysis by three different laboratories. Included in the tested cell lines were MCF-7 and MCF-7/ADR-RES. Utilizing restriction fragment length polymorphism (RFLP) testing, CellMark Diagnostics (Germantown, MD) concluded that their DNA fingerprinting data were consistent with each of the cell lines (MCF-7 and MCF-7/ADR-RES) having different donors. The other laboratories—American Type Culture Collection (Rockville, MD) used both RFLP and amplification fragment length polymorphism (AmpFLP) methods, and Children's Hospital of Michigan Cell Culture Laboratory (Detroit, MI) used the AmpFLP method—reached the same conclusions. Based on the reports from these DNA fingerprinting analyses, we have concluded that the preponderance of the information available suggests that the MCF-7/ADR-RES multidrug-resistant cell line that is included in the DTP screening program is not related to the MCF-7 cell line that is a part of the screening panel. **Thus, we have changed the nomenclature of the MCF-7/ADR-RES multidrug-resistant cell line.** The new designation of this cell line is **NCI/ADR-RES**. This nomenclature change will soon appear in all DTP databases, including the Worldwide Web. The DTP Web site address is: <http://dtp.nci.nih.gov>

Irrespective of its origin, this cell line has served as a valuable sentinel for compounds interacting with the multidrug-resistant mechanism (5).

DOMINIC A. SCUDIERO
ANNE MONKS
EDWARD A. SAUSVILLE

References

- (1) Monks A, Scudiero D, Skehan P, Shoemaker R, Paull K, Vistica D, et al. Feasibility of a high-flux anticancer drug screen using a diverse panel of cultured human tumor cell lines. *J Natl Cancer Inst* 1991; 83:757-66.
- (2) Stinson SF, Alley MC, Kopp WC, Fiebig HH, Mullendore LA, Pittman AF, et al. Morphological and immunocytochemical characteristics of human tumor cell lines for use in a disease-orientated anticancer drug screen. *Anticancer Res* 1992; 12:1035-53.
- (3) Wu L, Smythe AM, Stinson SF, Mullendore LA, Monks A, Scudiero DA, et al. Multidrug-resistant phenotype of disease-orientated panels of human tumor cell lines used for anticancer drug screening. *Cancer Res* 1992;52:3029-34.
- (4) Lee JS, Paull K, Alvarez M, Hose C, Monks A, Grever M, et al. Rhodamine efflux patterns predict P-glycoprotein substrates in the National Cancer Institute drug screen. *Mol Pharmacol* 1994; 46:627-38.
- (5) Alvarez M, Paull K, Monks A, Hose C, Lee JS, Weinstein J, et al. Generation of a drug resistance profile by quantitation of mdr-1/P-glycoprotein in the cell lines of the National Cancer Institute Anticancer Drug Screen. *J Clin Invest* 1995; 95:2205-14.

Notes

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NONHUMAN IN VITRO CELL LINES					
Designation	Species	Histologic Type	Tissue of Origin	Growth Medium	Remarks
CHO 1C T6	Hamster	Normal	Ovary	F12	
B16F ₁	Mouse	Melanoma	Ear (B16)	EMEM	From Fidler
B16F ₁₀	Mouse	Melanoma	Lung met.	EMEM	From Fidler; high lung met.
B16F ^{Lr6}	Mouse	Melanoma	Lung met.	EMEM	From Fidler; low lung met.
B16BL-6	Mouse	Melanoma	Bladder met.	EMEM	From Fidler; intermediate lung met.
BALB/c 3T3	Mouse	Normal	Embryo	Dulbecco	
C3HIOT 1/2	Mouse				No info
Colon 26	Mouse	Carcinoma	Colon	RPMI 1640	
EL-4	Mouse	Lymphoma			
FBL-3	Mouse	Leukemia			
Lewis Lung	Mouse	Carcinoma	Lung	EMEM	
L1210/MRI	Mouse	Leukemia	Ascites	Fischer's	
L5178Y(R)/MRI	Mouse	Leukemia	Ascites	Fischer's	
MPC-11	Mouse	Myeloma			
M5076	Mouse	Reticulum cell sarcoma		RPMI 1640	
P3X63	Mouse				No info
P388	Mouse	Leukemia	Ascites	RPMI 1640	
P388/ADR	Mouse	Leukemia	Ascites	RPMI 1640	
PAN 02	Mouse	Adenocarcinoma	Pancreas	RPMI 1640	
YAC	Mouse	Lymphoma		EMEM	
K-1735	Mouse	Melanoma		EMEM	
UV-2237	Mouse	Fibrosarcoma		EMEM	
MADB 106	Rat				
MADB 200	Rat				

YEAST STRAINS

YEAST STRAINS USED FOR NCI COMPOUND SCREENING		
SPY #	Relevant Mutation(s)	Complete Genotype
50636	<i>rad52</i>	<i>MATα rad52ΔURA3 erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 trp1 ura3 cyh2</i>
50644	none (wild-type control)	<i>MATα erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 ura3 cyh2</i>
50648	<i>rad50</i>	<i>MATα rad50Δkan^r ade2 ade3 leu2 ura3 trp1 cyh2</i>
50649	<i>mlh1 rad18</i>	<i>MATα mlh1ΔTRP1 rad18ΔLEU2 erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 ura3 cyh2 (trp1?)</i>
50650	<i>mgt1</i>	<i>MAT mtg1Δkan^r erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 trp1 ura3 cyh2</i>
50652	<i>rad50</i>	<i>MATα rad50Δkan^r erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 ura3 cyh2</i>
50654	<i>mec2-1</i>	<i>MATα mec2-1 erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 ura3 cyh2</i>
50740	<i>rad14</i>	<i>MATα rad14Δkan^r erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG::URA3::hisG ade2 ade3 leu2 ura3 cyh2</i>
50745	<i>sgs1 mgt1</i>	<i>MATα sgs1ΔLEU2 mgt1kan^r erg6ΔLEU2 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 ura3 cyh2</i>
50768	<i>GPDp-CLN2</i>	<i>MATα URA3-GPDp-CLN2 erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 trp1 ura3 cyh2</i>
50771	<i>GPDp-CLN2 rad14</i>	<i>MATα URA3-GPDp-CLN2 rad14Δkan^r erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 ura3 cyh2 trp1</i>
50779	<i>bub3</i>	<i>MATα bub3ΔURA3 erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 ura3 cyh2 trp1</i>
50780	none (wild-type control)	<i>MATα erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 trp1 ura3 cyh2</i>
50834	<i>mlh1</i>	<i>MATα mlh1ΔTRP1 erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 ura3 cyh2</i>
50835	<i>sgs1</i>	<i>MATα sgs1ΔLEU2 erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 ura3 cyh2</i>
50891	<i>rad18</i>	<i>MATα rad18ΔURA3 erg6ΔTRP1 pdr1ΔLEU2 pdr3ΔhisG ade2 ade3 leu2 trp1 ura3 cyh2</i>

Notes:

Store at -70°C to -80°C. To establish working stock: scrape frozen culture with a wooden applicator stick and apply sample to agar-containing media (vials should remain frozen).

All strains are derived from L. Hartwell laboratory strains in the A364a genetic background.

The *erg6 pdr1 pdr3* mutations in all strains serve to make yeast more sensitive to a variety of compounds.

The allele present at the *TRP1* locus is unknown for SPY50649 (strain is phenotypically Trp⁺ by virtue of *TRP1* at the *MLH1* locus).

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8 Lym-108(1)	Lymphatic Leukemia	Rat	55
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CA36/Ara C (NSC-63878)	Colon Adenocarcinoma	Mouse	31
CA51	Colon Adenocarcinoma	Mouse	31
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Glioma 261	Glioma	Mouse	33
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L1210/C95	Lymphoid Leukemia	Mouse	34
L1210/cis-DDP (NSC-119875)	Lymphoid Leukemia	Mouse	34
L1210/CTX (NSC-26271)	Lymphoid Leukemia	Mouse	34
L1210/DF8 (NSC-29630)	Lymphoid Leukemia	Mouse	34
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Morris Hepatomas	Hepatoma	Rat	51
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MPC-25	Plasmacytoma	Mouse	38
MPC-26	Plasmacytoma	Mouse	38
MPC-31	Plasmacytoma	Mouse	38
MPC-36	Plasmacytoma	Mouse	38
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MRI-H-121	Kidney, Carcinoma	Human	17
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MRI-H-130	Cervix, Squamous Cell Carcinoma	Human	16
MRI-H-147	Endometrium, Carcinoma, Müllerian Duct	Human	17
MRI-H-1579	Prostate Adenocarcinoma	Human	18
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MRI-H-1834	Ovarian Carcinoma	Human	17
MRI-H-186	Cervix, Invasive, Large Cell, Nonkeratinizing, Squamous Cell Carcinoma	Human	16
MRI-H-187	Melanoma, Epitheloid, Melanotic	Human	16
MRI-H-194	Colon, Adenocarcinoma	Human	15
MRI-H-196	Cervix, Poorly Differentiated Squamous Cell Carcinoma	Human	16
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MRI-H-266	Lung, Poorly Differentiated Carcinoma	Human	15
MRI-H-273	Ovarian Carcinoma	Human	17
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MST	Mast Cell	Mouse	38
MT/W449	Mammary Adenocarcinoma	Rat	51
MT/W9	Mammary Adenocarcinoma	Rat	51
MtT	Anterior Pituitary	Rat	51
Murphy-Sturm Lymphosarcoma (MSL)	Lymphosarcoma	Rat	51
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MX-2	Mammary Carcinoma	Human	14
MXT	Mammary Ductal	Mouse	38
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NC-37	Normal	Human <i>In Vitro</i> Cell Lines	59
NCI/ADR-RES	Adenocarcinoma	Human <i>In Vitro</i> Cell Lines	59
NCI-H125	Adenosquamous Carcinoma	Human <i>In Vitro</i> Cell Lines	59
NCI-H226	Squamous Cell	Human <i>In Vitro</i> Cell Lines	59
NCI-H23	Adenocarcinoma	Human <i>In Vitro</i> Cell Lines	59
NCI-H292	Adenosquamous Carcinoma	Human <i>In Vitro</i> Cell Lines	59
NCI-H322M	Adenocarcinoma	Human <i>In Vitro</i> Cell Lines	59
NCI-H358M	Bronchiolo-Alveolar Carcinoma	Human <i>In Vitro</i> Cell Lines	59
NCI-H460	Large Cell	Human <i>In Vitro</i> Cell Lines	59
NCI-H522	Adenocarcinoma	Human <i>In Vitro</i> Cell Lines	59
NCI-H69	Small Cell Carcinoma	Human <i>In Vitro</i> Cell Lines	59
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Nettesheim Lung	Squamous Cell Lung Carcinoma	Mouse	38
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Nitrosamine-Induced Hepatoma	Hepatoma	Guinea Pig	22
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Novikoff Hepatoma	Hepatoma	Rat	51
NS104	Rhabdomyosarcoma	Rat	51
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P1798/CR-JS	Lymphoma	Mouse	40
P1798/CS-JS	Lymphoma	Mouse	40
P288	Lymphocytic Leukemia	Mouse	39
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P388	Lymphocytic Leukemia	Mouse	39
P388	Leukemia	NonHuman <i>In Vitro</i> Cell Lines	66
P388/5-Azacytidine (NSC-102816)	Lymphocytic Leukemia	Mouse	39
P388/5FU (NSC-19893)	Lymphocytic Leukemia	Mouse	39
P388/Acivicin (NSC-163501)	Lymphocytic Leukemia	Mouse	39
P388/ActinomycinD (NSC-3053)	Lymphocytic Leukemia	Mouse	39
P388/ADR (NSC 123127)	Lymphocytic Leukemia	Mouse	39
P388/Amsacrine (NSC-249992)	Lymphocytic Leukemia	Mouse	39
P388/Ara-A + 2'dcF (NSC-404241 + NSC-218321)	Lymphocytic Leukemia	Mouse	39
P388/Ara-C (NSC-63878)	Lymphocytic Leukemia	Mouse	39
P388/BCNU (NSC-409962)	Lymphocytic Leukemia	Mouse	39
P388/CPA (NSC-26271)	Lymphocytic Leukemia	Mouse	39
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P388/DON (NSC-7365)	Lymphocytic Leukemia	Mouse	39
P388/L-Alanosine (NSC-153353)	Lymphocytic Leukemia	Mouse	39
P388/L-PAM (NSC-8806)	Lymphocytic Leukemia	Mouse	39
P388/Mitoxantrone (NSC-301739)	Lymphocytic Leukemia	Mouse	39
P388/MTX (NSC-740)	Lymphocytic Leukemia	Mouse	39
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P815 (Hageman Mastocytoma)	Mast Cell Leukemia	Mouse	40
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Pan #1 (Fortner)	Pancreatic Duct Adenocarcinoma	Hamster	20
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PAN 02	Adenocarcinoma	NonHuman <i>In Vitro</i> Cell Lines	66
PAN 03	Pancreas	Mouse	40
Papillary Carcinoma		Mouse	
PC-3	Carcinoma	Human <i>In Vitro</i> Cell Lines	60
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PR1C1T5/NSC-45388	Lymphoid Leukemia	Mouse	40
PR1SE1T5	Lymphoid Leukemia	Mouse	40
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RPMI-7951	Melanoma	Human <i>In Vitro</i> Cell Lines	60
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RXF 393 (available only to DTP, NCI)	Renal Cell Carcinoma	Human <i>In Vitro</i> Cell Lines	60
RXF 631	Renal Cell Carcinoma	Human <i>In Vitro</i> Cell Lines	60
S180	Pleomorphic Sarcoma	Jackson Lab Mouse	45
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S37	Pleomorphic Cell	Mouse	40
S37	Pleomorphic Sarcoma	Jackson Lab Mouse	45
S91	Melanoma (Melanotic)	Jackson Lab Mouse	44
Sa 1	Spindle Cell Carcinoma	Mouse	41
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Sal	Spindle-Cell Sarcoma	Jackson Lab Mouse	45
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Sarcoma		Mouse	
SB #1 (Fortner)	Small Bowel Adenocarcinoma	Hamster	20
SF 295	Glioblastoma	Human	18
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SF-295	Glioblastoma	Human <i>In Vitro</i> Cell Lines	60
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SK-MEL-5	Melanoma	Human <i>In Vitro</i> Cell Lines	60
SK-MES-1	Squamous Cell Carcinoma	Human <i>In Vitro</i> Cell Lines	60
SK-OV-3	Adenocarcinoma	Human <i>In Vitro</i> Cell Lines	60
SMT-2A	Mammary Carcinoma	Rat	52
SN12A1	Renal Cell Carcinoma	Human <i>In Vitro</i> Cell Lines	60
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SN12L1	Renal Cell Carcinoma	Human <i>In Vitro</i> Cell Lines	60
SN21S1	Renal Cell Carcinoma	Human <i>In Vitro</i> Cell Lines	60
SN12K1	Renal Cell Carcinoma	Human <i>In Vitro</i> Cell Lines	60
SNB-19	Glioblastoma	Human <i>In Vitro</i> Cell Lines	60
SNB-7	Glioblastoma	Human <i>In Vitro</i> Cell Lines	60
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Spontaneous DBA/2 Mammary	Mammary Adenocarcinoma	Mouse	41
SR	Lymphoma	Human <i>In Vitro</i> Cell Lines	60
SV-122-TR4		Mouse	41
SW-613	Mammary Carcinoma	Human	14
SW-620		Human <i>In Vitro</i> Cell Lines	60
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T1699	Mammary Adenocarcinoma	Jackson Lab Mouse	44
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WIDR (derivative of HT-29)	Adenocarcinoma	Human <i>In Vitro</i> Cell Lines	61
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XF 498	Glioblastoma	Human <i>In Vitro</i> Cell Lines	61
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